Synovial tissue macrophages: friend or foe?

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**ABSTRACT**

Healthy synovial tissue includes a lining layer of synovial fibroblasts and macrophages. The influx of leucocytes during active rheumatoid arthritis (RA) includes monocytes that differentiate locally into proinflammatory macrophages, and these produce pathogenic tumour necrosis factor. During sustained remission, the synovial tissue macrophage numbers recede to normal. The constitutive presence of tissue macrophages in the lining layer of the synovial membrane in healthy donors and in patients with RA during remission suggests that this macrophage population may have a role in maintaining and reinstating synovial tissue homeostasis respectively. Recent appreciation of the different origins and functions of tissue-resident compared with monocyte-derived macrophages has improved the understanding of their relative involvement in organ homeostasis in mouse models of disease. In this review, informed by mouse models and human data, we describe the presence of different functional subpopulations of human synovial tissue macrophages and discuss their distinct contribution to joint homeostasis and chronic inflammation in RA.

**SYNOVIAL TISSUE HOMEOSTASIS**

Synovial joints link the musculoskeletal system, and the joint synovium facilitates movement. Bones are linked by the joint capsule and collagenous ligaments, and the bone surface is covered with articular cartilage that absorbs impact. The joint capsule consists of an outer fibrous membrane containing ligaments and proprioceptive sensory nerves that regulate the posture and motion of the joint and an inner synovial membrane that produces synovial fluid that lubricates the joint during movement and nourishes avascular cartilage. In healthy people, this complex joint structure is self-regulated by homeostatic mechanisms to maintain well-functioning articulation, and the synovial membrane is vital for maintaining joint homeostasis.1 In rheumatoid arthritis (RA), this normal joint structure is progressively compromised due to inflammation of the synovial membrane that fails to resolve, and this ultimately leads to loss of joint function.

The synovial membrane is a highly specialised, multifunctional structure consisting of two layers: the first is a thin (eg, <2 mm in radiocarpal joints by ultrasound imaging)2 but highly cellular lining layer composed of two cell types: synovial fibroblasts and synovial macrophages,1 and the second is a supporting sublining layer containing loose connective tissue with sublining fibroblasts, a rich network of sympathetic and sensor nerves and blood and lymphatic vasculature that provides oxygen/nutrients and immune-drainage.1 Synovial fibroblasts provide the extracellular matrix (ECM) that supports the structure of the synovium and secretes hyaluronic acid and lubricin into the synovial fluid.1 Synovial tissue macrophages (STM) are constitutively resident in healthy synovium. Their tissue-specific function remains unresolved, but this likely includes sentinel joint homeostasis. Other immune cells (lymphocytes, mast cells and dendritic cells) are scarce in the normal synovium and localise mainly in perivascular areas of the sublining layer.3

In patients with RA, the synovial membrane becomes hypertrophic (eg, 2–5 mm in radiocarpal joints)2 due to synovial fibroblast proliferation, increased blood/lymphatic vasculature and an inflammatory influx of immune cells from the circulation. The products of these activated cells lead to destruction of cartilage and bone, pain and loss of joint function.1 These changes appear consequential to aberrant homeostatic mechanisms
that fail to resolve inflammatory synovitis. Understanding synovial homeostasis may offer the prospect of new therapeutic strategies for RA.

SYNOVIAL TISSUE INFLAMMATION AND RESOLUTION IN RA

RA is the most common inflammatory arthropathy, affecting approximately 0.4 million people in the UK, causing morbidity and reduced quality of life and represents a substantial socioeconomic and healthcare burden. An interplay between genetics (>100 loci), environmental factors (eg, cigarette smoking and occupational exposure to silica) and alterations in the mucosal microbiome can contribute to the development of RA. Clinical RA is preceded by an asymptomatic preclinical breach of tolerance to citrullinated self-peptides, activation of autoreactive T cells and production of anticitrullinated peptide-specific antibodies (ACPAs). Joint injury or other poorly defined events can then lead to activation of the synovial tissue vasculature permitting localisation of ACPA immune complexes in the synovium. These complexes activate synovial tissue fibroblasts and bone cells (osteoclasts) and perpetuate the influx of leukocytes, for example, neutrophils, monocytes, Th1/Th17 and B cells. These cells are activated by local stimuli (eg, endogenous Toll-like receptors (TLR) ligands, proinflammatory cytokines and hypoxia), further modulated by genetic and epigenetic modifications of gene expression to propagate the synovitis. Depending on the dominant cellular component and distinct molecular pathogenic pathways, RA synovitis can be characterised as myeloid (macrophage rich), lymphoid (containing T/B cell follicles) and fibroid (leucocyte low) (comprehensive reviews on mechanisms and type of synovitis). Emerging data suggest that this underlying cellular heterogeneity in RA may influence the clinical outcome to therapies targeting different biological pathways.

Recent advances in RA therapies mainly target the mediators of inflammation (eg, tumour necrosis factor (TNF), interleukin (IL)-6R and janus kinase (JAKs)), or block activators of inflammation (eg, tumour necrosis factor (TNF), interleukin (IL)-6R and janus kinase (JAKs)), or block inhibitors of inflammation (eg, tumour necrosis factor (TNF), interleukin (IL)-6R and janus kinase (JAKs)). RA synovitis can be characterised as myeloid (macrophage rich), lymphoid (containing T/B cell follicles) and fibroid (leucocyte low) (comprehensive reviews on mechanisms and type of synovitis). Emerging data suggest that this underlying cellular heterogeneity in RA may influence the clinical outcome to therapies targeting different biological pathways.

STM SUBPOPULATIONS

The functions of macrophages in joint synovial tissue of healthy subjects are poorly described. In RA, most knowledge of synovial myeloid cells is derived from studies of monocytes from RA synovial fluid that contribute to local production of inflammatory and joint-degrading mediators. However, monocytes are not present in the synovial fluid from healthy subjects and from patients with RA in remission. STMs are the most common resident immune cells in the healthy synovial membrane, and dendritic cells and lymphocytes are scarce. In active RA with myeloid and lymphoid synovitis, the synovial membrane is leucocyte-rich, including an increased number of proinflammatory macrophages that likely differentiate locally from blood monocytes attracted to synovial tissue and fluid by local chemokines, and these macrophages are the main producers of pathogenic TNF. In remission, most of the synovial inflammation resolves, but STMs in the lining layer persist. It would be important to establish if these macrophages have the same phenotype as those in healthy synovium. The constitutive presence of tissue macrophages in healthy synovial tissue and the sustained presence in patients with RA during remission suggests that these subpopulations of STMs may have a role in synovial homeostasis.

THE CONCEPT OF TISSUE-RESIDENT AND MONOCYTE-DERIVED MACRHOPHAGES

Mouse models show that most healthy organs contain tissue-resident macrophages that originate from prenatal precursors and are essential for maintaining immune homeostasis and intact organ function. In response to inflammation, a different tissue macrophage population differentiates from monocytes that migrate transiently from the circulation. The relative roles of these macrophage populations determine chronicity or resolution of inflammation.

The ontogeny and precise role of human STMs in joint homeostasis are unknown. Animal studies suggest that mouse resident STMs are of prenatal origin and can proliferate locally in the tissue, whereas synovial tissue proinflammatory macrophages differentiate from either of two subpopulations of blood monocytes (Ly6C(−) and Ly6C(+) that traffic into the inflamed synovium dependent on the nature of the trigger of the joint inflammation. The Ly6C(−) patrolling monocytes differentiate into proinflammatory macrophages that mediated joint pathology in an antibody transfer model of arthritis, while Ly6C(+) classical monocytes gave rise to synovial macrophages that drive inflammation in adjuvant-induced and
The role of synovial tissue macrophages in arthritis. (A) The mouse synovium contains major histocompatibility complex (MHC) class II negative (MHC-II−) tissue-resident macrophages that originate from prenatal precursors and have the capacity to proliferate. On induction of experimental arthritis, there is an influx of monocytes (Ly6C+ or Ly6C−, depending of the nature of the induction) that differentiate into MHC class II positive (+) proinflammatory macrophages that mediate joint pathology. Resolution of arthritis is associated with a phenotypic change of the monocyte-derived macrophages from proinflammatory to anti-inflammatory, along with the contribution of tissue-resident macrophages. (B) In humans, the synovial membrane of healthy subjects and of patients with RA in sustained remission contain macrophages in the lining layer. Their origin and function are unknown. In patients with active arthritis, the CD14+ subpopulation of infiltrating monocytes contributes to the increased MHC-II+ proinflammatory macrophage pool in the inflamed synovium. Proliferative macrophages of patients with arthritis produce a broad range of inflammatory mediators, and their phenotype is maintained by the miR-155/SHIP-1 pathway. RA, rheumatoid arthritis; SHIP-1, phosphatidylinositol-3,4,5-trisphosphate 5-phosphatase 1; TLR, Toll-like receptors.

antigen-induced arthritis. It is likely that the difference in chemokines induced by immune complexes or adjuvant in these models were responsible for attracting different populations of monocytes into the joints. In vivo tracking of labelled blood monocytes (CD14+ population that resembles mouse Ly6C+) in patients with RA has confirmed their migration into the inflamed synovium where they add to the tissue macrophage pool during inflammation. The traffic of other monocyte subpopulations into and within synovial tissue remains to be confirmed (figure 1).

The distinct functions of mouse tissue-resident and monocyte-derived macrophage populations in animal models of disease have improved our understanding of organ homeostasis. Tissue-resident macrophages can be self-renewing, controlled by the transcription factor Gata6, and maintain tissue immune homeostasis, whereas monocyte-derived macrophages predominate and drive inflammation in chronic disease models.

Successful resolution of inflammation and reinstatement of tissue homeostasis requires differentiation of these proinflammatory macrophages towards an anti-inflammatory phenotype, and the activation/expansion of resident tissue macrophages that synergise for tissue homeostasis by production of anti-inflammatory/immune regulatory mediators (eg, resolvins, IL-10 and transforming growth factor beta (TGFβ)). In addition, tissue-resident macrophages contribute to tissue remodelling by removing apoptotic cells (effecrocytosis), metabolic products and damaged tissue components, and by reinstating tissue spatial organisation, for example, pigment cell distribution in zebrafish skin. In addition to the shared immune regulatory functions, mouse tissue-resident macrophages show tissue-specific homeostatic properties that are determined by the physiological needs of their specific environment. Tissue-resident macrophage differentiation and function is governed by tissue-specific cues and ‘on demand’ signals. Examples of these have been shown in an elegant study by van der Laar and colleagues in which either yolk-sac macrophages, fetal liver macrophages or bone marrow monocytes, on transfer into empty lung alveolar niches, indistinguishably differentiated into alveolar macrophages with the characteristic local functional regulation of lung surfactants. Transfer of mature macrophages from other tissues could not replicate this differentiation, thus confirming that the plasticity of macrophage maturation is optimal at the precursor stage and that tissue-specific cues determine local macrophage specialisation.
Figure 2  What is the tissue-specific function of synovial-resident macrophages? Tissue-specific functions of resident-tissue macrophages are induced by local cues. Their function in the synovium is unknown, but we can speculate by reference to other tissues. In the lung, for example, the differentiation of alveolar macrophages is driven by lung epithelial cell-derived GM-CSF, and recycling of surfactant is regulated by transcription factor PPARγ. Deficiency of alveolar macrophages, GM-CSF or PPARγ leads to pulmonary proteinosis. Spleen red pulp macrophages specialised in iron-recycling. This function is induced by heme derived from erythrocyte degradation and executed by Bach2 and SPIC. Selective deficiency of red pulp macrophages (deficiency of SPIC) leads to aberrant iron metabolism. Osteoclasts are bone macrophages that specialise in bone degradation. Their functional programme is induced by RANKL and executed by multiple transcription factors, including NFATC1. Deficiency in functional osteoclasts (RANKL−/− or M-CSF−/−) leads to an increase in bone mass (osteopetrosis). Speculatively, in healthy joints, synoval tissue macrophages may specialise in recycling lubricin, the lubricating components of synovial fluid and in providing regulatory factors for cartilage and bone turnover. BACH2, transcription regulator protein BACH2; FLS, fibroblast-like synoviocytes; GM-CSF, granulocyte macrophage colony-stimulating factor; M-CSF, macrophage colony-stimulating factor; MQ, macrophages; NFATC1, nuclear factor of activated T cells, cytoplasmic 1; PPARγ, peroxisome proliferator-activated receptor gamma; RANKL, receptor activator of nuclear factor kappa-B ligand; SPIC, transcription factor Spi-C.

These tissue-specific cues include a combination of cell contact and soluble factors. These have not been delineated in the synovium, but equivalent examples have been described in other tissues (figure 2), for example, direct contact with neurons is required for macrophage precursors to develop into microglia. The tissue-soluble factors include cytokines and tissue breakdown products. The differentiation of alveolar macrophages requires lung epithelial cell-derived granulocyte macrophage colony-stimulating factor (GM-CSF) and the induction of the transcription factor peroxisome proliferator-activated receptor gamma (PPARγ) that contributes to tissue-specific local function by regulating lung surfactant. The local differentiation of spleen red pulp macrophages requires the transcription factor BACH1 that recognises heme from erythrocyte degradation and induces their specialised iron-recycling function. Retinoic acid produced by the omentum is key in the differentiation of peritoneal macrophages that function to control IgA production by B-1 cells, and receptor activator of nuclear factor kappa-B ligand (RANKL) expressed in bone osteoblasts drives the differentiation of macrophages (osteoclasts) with bone-resorbing function. A combination of these tissue-specific cues and transient ‘on-demand’ signals, (eg, tissue injury creates signals to remove apoptotic inflammatory cells) induces programmes that are common for all tissue macrophages or programmes that are specific for a particular tissue. These programmes enable cells to perform general immune homeostatic functions, for example, Mer tyrosine protein kinase receptor (MerTK)-mediated effrocytosis and production of anti-inflammatory resolvins, or tissue-specific functions, for example, iron recycling by the spleen macrophages. These tissue-specific transcriptomic programmes are coordinated by specific transcription factors, for example, SPIC in spleen red pulp, LXRa in spleen marginal zone, PPARγ2 and Bach2 in alveolar macrophages, GATA6 in peritoneal macrophages and NR1A in thymic macrophages. These cooperate with macrophage lineage master regulator transcription factors such as PU.1 induced by macrophage colony-stimulating factor (M-CSF)/IL-34 that maintain universal macrophage-specific gene expression. Alveolar macrophage deficiency in humans and mice results in pulmonary alveolar proteinosis caused by the progressive accumulation of surfactant, and mice deficient in functional spleen red pulp macrophages develop a disorder of iron homeostasis. These examples illustrate the critical role of tissue-resident macrophages in the maintenance of tissue homeostasis (figure 2).

HUMAN HOMEOSTATIC STMS: FRIEND

The diversity of human STMs in health and disease is poorly described, while their tissue-specific functions are unknown. In healthy joints, STMs are located on the surface of the synovial membrane and are in contact with synovial fluid, suggesting that they may specialise in recycling the lubricating components of synovial fluid (analogous to alveolar macrophage recycling surfactant) and in providing regulatory factors for cartilage and bone turnover. Similar to other tissue-resident macrophages, they may also remove cell debris and pathogens to prevent sterile and septic inflammation. We review here the evidence for the potential homeostatic role of human synovial tissue-resident macrophages. Most of these data are historical and/or based on immunohistochemistry and is therefore of limited functional and molecular resolution.
Early studies described three subtypes of STMs distinguishable by staining with 25F9, 27E10 and RM3/1 antibody clones. Their tissue distribution differed depending on whether the synovial tissue was normal, inflamed or resolving in studies on septic arthritis, suggesting that they had distinct functions. In patients with osteoarthritis the 25F9+ macrophages predominated in the non-inflammatory synovium and in the non-inflamed areas in the synovium of septic arthritis, whereas macrophages in the inflamed areas in the synovium of septic arthritis were 27E10+ and TNF antibody therapy after which they are removed by rapid efflux. RM3/1 (CD163) macrophages, typically found in healthy joints, are also present in RA synovium. They were found to be localised far from IFNγ positive T cells and their numbers were unaffected by anti-TNF therapy, suggesting that they have distinct functions. Historical ex vivo functional studies demonstrated that total macrophages isolated from active RA synovial tissue spontaneously released proinflammatory mediators (eg, TNF, IL-1β, IL-6, CXCL5, CXCL7 and CCL2) and are able to stimulate autologous T cells. Staining of these macrophages with the antibody recognising CD13, expressed on bone marrow-derived myeloid cells, alluded to the presence of at least two functionally distinct subpopulations that differed in their angiogenic properties. Only macrophages that were CD13 positive promoted angiogenesis, implying a role in vascularisation and hyperplasia of the RA synovium.

The activation of proinflammatory macrophages in humans and mice is governed by the transcription factors NFXb, IRF5 and STAT1/5 and is maintained by microRNA-155. We showed that increased miR-155 prevented the translation of its mRNA epigenetic target SHIP-1, and thereby removed its negative feedback control of inflammatory TLR and PI3K signalling, thus permitting sustained production of cytokines and chemokines by these macrophages. Most RA STMs express high levels of miR-155 and commensurately low levels of SHIP-1, which is in sharp contrast to macrophages from non-inflamed synovium where the majority lack miR-155 and have high expression of SHIP-1 (figure 3).

Together these data provide evidence for different macrophage subpopulations in human synovium with potentially homeostatic or inflammatory properties. However, the prospect of therapeutic translation of the homeostatic potential of STM subpopulations requires more precise understanding of their phenotypic heterogeneity and function. The rapid progress in using synovial tissue biopsy to study disease pathogenesis and the current advances in a single cell sequencing could help to reveal their biology.

**HUMAN INFLAMMATORY STMS: FOE**

In active RA, fuelled by an influx of blood monocytes, the total number of STMs increased in all types of synovitis, particularly in the sublining layer of myeloid and lymphoid types. They are predominantly proinflammatory and produce CXCL4 and CXCL7 (chemokines recruiting neutrophils and blood monocytes) particularly in early RA, and TNF and other pro-inflammatory cytokines (IL-15, IL-1β, IL-6, GM-CSF and TGFβ) throughout disease progression. These macrophages are 27E10+, which recognises an epitope on the alarmins S100A8/9. These S100A8/9 macrophages are susceptible to anti-TNF therapy after which they are removed by rapid efflux. RM3/1 (CD163) macrophages, typically found in healthy joints, are also present in RA synovium. They were found to be localised far from IFNγ positive T cells and their numbers were unaffected by anti-TNF therapy, suggesting that they have distinct functions. Historical ex vivo functional studies demonstrated that total macrophages isolated from active RA synovial tissue spontaneously released proinflammatory mediators (eg, TNF, IL-1β, IL-6, CXCL5, CXCL7 and CCL2) and are able to stimulate autologous T cells. Staining of these macrophages with the antibody recognising CD13, expressed on bone marrow-derived myeloid cells, alluded to the presence of at least two functionally distinct subpopulations that differed in their angiogenic properties. Only macrophages that were CD13 positive promoted angiogenesis, implying a role in vascularisation and hyperplasia of the RA synovium.

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**POTENTIAL TISSUE-SPECIFIC ‘CUES’ AND ‘ON DEMAND’ DETERMINANTS OF SYNOVIAL TISSUE MACROPHAGE HOMEOSTATIC AND INFLAMMATORY FUNCTIONS**

Okabe and Medzhitov proposed a model in which the tissue-resident macrophage phenotype is determined by core transcriptomic modules induced, for example, by M-CSF/PU.1 that are common for all macrophages. Thereafter, tissue-specific transcriptomic modules are induced by tissue ‘cues’ (eg, GM-CSF in alveolar macrophages) that determine tissue-specific function (eg, recycling of surfactant in the lung) to maintain organ-specific homeostasis. Changes in the local environment, for example, infection or tissue injury, induce ‘on demand’ modules to eliminate the stressor, for example, chemokine production, to initiate the recruitment of neutrophils and monocytes that become a source of

What are the effector pathways, transcriptional regulators and activators of synovial tissue macrophages in healthy donors and in patients with RA in remission? The constitutive presence of tissue macrophages in healthy synovial tissue and the sustained presence of some synovial tissue macrophages in patients with RA during remission suggests that these subpopulations have a role in maintaining and reinstating synovial homeostasis. However, little is known about the effector pathways and stimuli and transcription factors that execute their function. Synovial tissue macrophages in healthy subjects express the scavenger receptor CD163, suggesting that they are strongly phagocytic. They also express MHC-II, IL-1R-antagonist and the inhibitor of bone degradation, osteoprotegerin (OPG), and they are negative for proinflammatory cytokines (eg, TNF and IL1β) and the bone resorption cytokine RANKL, suggesting a joint protective function against inflammation and damage. In contrast, the phenotype of proinflammatory synovial tissue macrophages is well described. These cells are MHC-II positive and produce a broad range of inflammatory mediators (eg, TNF, IL1β, IL-6, IL-23 and S100A8/9) that drive local and systemic pathologies in RA. Their activation is sustained by a variety of local stimuli that include endogenous TLR ligands, immune complexes, oxidised lipids, hypoxia and integrin-mediated contact with synovial fibroblasts and T cells. This proinflammatory programme is executed by NFκB, IRF5, STAT1/5 and HIF1α and is maintained by microRNA-155. FcγR, Fc gamma receptor; GM-CSFR, granulocyte macrophage colony-stimulating factor receptor; IL, interleukin; IRF, interferon regulatory factor; LXRα, liver X receptor alpha; NFκB, nuclear factor kappa-light-chain-enhancer of activated B cells; RA, rheumatoid arthritis; STAT, signal transducer and activator of transcription; TLR, Toll-like receptors; TNF, tumour necrosis factor.

inflammatory macrophages or efferocytosis to facilitate the reinstatement of tissue homeostasis at the later stages of inflammation.

The tissue-specific cues determining the phenotype and transcriptional programme of synovial tissue-resident macrophages in health and those present in patients with RA in remission are unknown. Local differentiation of microglia requires direct contact of primitive macrophage precursors with neurons, therefore by analogy, the interaction of macrophage precursors with lining synovial fibroblasts that undergo biomechanical stretch may trigger their local differentiation into synovial tissue-resident macrophages. The characteristic ECM produced by synovial fibroblasts (eg, lubricin and hyaluronic acids), by analogy to the profound impact of ECM produced by lung fibroblasts on the transcriptome of recruited myeloid cells, could affect macrophage differentiation and survival by binding to macrophage CD14 or TLR2. Uncovering the transcriptomic programme, molecular regulators and cues that drive synovial tissue-resident macrophage function will aid the understanding of mechanisms of joint homeostasis and provide novel therapeutic approaches to reinstate joint immune homeostasis in patients with arthritis.

In the tissue environment of the RA synovium, tissue-specific cues are modulated by tissue injury and proinflammatory ‘on-demand’ signals, which mobilise monocytes and instruct their differentiation to monocyte-derived pro-inflammatory macrophages. It is unknown whether distinct human tissue-resident and monocyte-derived synovial tissue macrophage subpopulations would respond differently to the synovial tissue-specific and ‘on demand’ proinflammatory cues (figure 4). In mouse tissues, recruitment of blood monocytes in response to ‘on-demand’ signal is often concomitant with a loss of tissue-resident macrophages due to emigration
Rheumatoid arthritis

**Figure 4** Putative contribution of tissue-specific (signal 1) and the local polarisation environment (signal 2) to the phenotype of synovial tissue macrophages in homeostasis and inflammation. Tissue-specific cues (signal 1), for example, from stretched synovial fibroblasts (FLS) to macrophage precursors may induce the synovial tissue macrophage programme that maintains synovial homeostasis. In the inflamed RA synovium, tissue-specific cues are modulated by proinflammatory on-demand signals (signal 2; eg, provided by epigenetically changed FLS, leucocytes and hypoxia). Most understanding is based on experimental models, and it is unknown whether distinct human synovial tissue macrophage subpopulations would respond differently to the synovial tissue-specific and ‘on-demand’ proinflammatory cues. Dissecting these pathways will improve our understanding of the mechanisms of successful versus failed joint homeostasis. MC, mast cells; MQ, macrophages; Neu, neutrophils; TF, transcription factor; Th17/Th1, T helper 1 and 17.

or death. In the synovium of active patients with RA, both S100A8/9 and CD163+ macrophages are present, although the latter in much lower numbers. Similarly, in mouse models of arthritis, tissue-resident macrophages were present at the peak of inflammation, along with monocyte-derived macrophages, although outnumbered and with distinct transcriptomic profiles. For example, monocyte-derived macrophages expressed high levels of proinflammatory mediators, for example, IL-1β, IL-12, CD80 and CD86, compared with resident macrophages, while the latter upregulated efferocytosis receptors. This suggests that, at least in the mouse, tissue-resident and monocyte-derived macrophages may respond differently to ‘on demand’ signals. In RA synovium, the phenotype of monocyte-derived macrophages is shaped by the interaction with synovial fibroblasts that are epigenetically reprogrammed to produce a wide range of mediators, for example, GM-CSF that potentiates the proinflammatory activation of macrophages. In addition, danger-associated molecular pattern molecules (DAMPs) released in the damage joint, for example, tenasin-C, bind to TLR4 and trigger proinflammatory cytokine production by macrophages. ACPA produced locally in lymphoid type of synovitis forms immune complexes with citrullinated fibrinogen and induces TNF production via synergistic binding to FcγR and TLR4. Oxyesters, enriched in RA synovial fluid, bind to LXRα in synovial macrophages and enhance DAMPs-induced TNF, IL-6 and IL-1β production. Memory T cells, recruited and expanded in synovial fluid by cytokines (eg, IL-15, IL-6 and TNF), trigger substantial production of TNF by macrophages on direct integrin-mediated contact, while IFNγ released by RA citrullinated peptide autoreactive T cells enhances the ability of macrophages to present antigen and thereby their potential to activate autoreactive memory T cells. The hypertrophic synovium creates a hypoxic environment, and recruited macrophages adapt by changing their metabolism and phenotype. This response, mediated by hypoxia inducible transcription factor 1 (HIF-1α), leads to an increased production of proinflammatory cytokines and can redirect differentiation towards bone-resorbing osteoclasts. For example, HIF-1α abrogates the mTORC1/IL-10-induced negative feedback mechanism of macrophage activation while stimulating IL-1β production. In addition, hypoxia decreases the expression of COMMD1 (an inhibitor of macrophage to osteoclast differentiation) and can thereby facilitate synovial macrophage-driven pathogenic bone resorption. Many of these signals increase the expression of miR-155, which locks the macrophages in a proinflammatory state. In mouse models of arthritis, at some time-point, there is a phenotypic change of monocyte-derived macrophages from proinflammatory to anti-inflammatory, which is induced by an as-yet unidentified signal.
This phenotypic change in monocyte-derived macrophages and the functions of tissue-resident macrophages are required for the resolution of arthritis.  

In RA, the role of recruited synovial macrophages in mediating remission of disease is uncertain. The observation that alleviation of disease with anti-TNF therapy leads to a decrease of joint-infiltrating S100A8/9+ macrophages, presumably by enhanced efflux of these cells from the synovium, while the number of potential resident-tissue (CD163+) macrophages remained unchanged would argue against an active role of monocyte-derived macrophages in the resolution of RA.

**SUMMARY**

There is evidence for the presence of different subpopulations of synovial tissue macrophages with potentially distinct homeostatic versus inflammatory functions. While synovial macrophage inflammatory pathways are relatively well described and therapeutically targeted, little is known about the synovial tissue macrophages constitutively present in healthy subjects and persistent in patients with RA during sustained remission. Understanding the biology of these cells could help to reveal the mechanisms of synovial immune homeostasis and could offer the prospect of novel strategies for treatment of joint diseases.

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