





ORIGINAL RESEARCH

Dysregulated long non-coding RNA in
Sjögren's disease impacts both interferon
and adaptive immune responses

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ABSTRACT

Objective Sjögren's disease (SjD) is an autoimmune disease characterised by inflammatory destruction of exocrine glands. Patients with autoantibodies to Ro/SSA (SjD^{Ro+}) exhibit more severe disease. Long non-coding RNAs (lncRNAs) are a functionally diverse class of non-protein-coding RNAs whose role in autoimmune disease pathology has not been well characterised.

Methods Whole blood RNA-sequencing (RNA-seq) was performed on SjD cases (n=23 Ro/SSA negative (SjD^{Ro-}); n=27 Ro/SSA positive (SjD^{Ro+}) and healthy controls (HCs; n=27). Bioinformatics and pathway analyses of differentially expressed (DE) transcripts (log₂ fold change ≥2 or ≤-0.5; p_{adj} <0.05) were used to predict lncRNA function. *LINC01871* was characterised by RNA-seq analyses of HSB-2 cells with CRISPR-targeted *LINC01871* deletion (*LINC01871*^{-/-}) and in vitro stimulation assays.

Results Whole blood RNA-seq revealed autoantibody-specific transcription profiles and disproportionate downregulation of DE transcripts in SjD cases relative to HCs. Sixteen DE lncRNAs exhibited correlated expression with the interferon (IFN)-regulated gene, *RSAD2*, in SjD^{Ro+} (r≥0.65 or ≤-0.6); four antisense lncRNAs exhibited IFN-regulated expression in immune cell lines. *LINC01871* was upregulated in all SjD cases. RNA-seq and pathway analyses of *LINC01871*^{-/-} cells implicated roles in cytotoxic function, differentiation and IFN γ induction. *LINC01871* was induced by IFN γ in a myeloid cell line and regulated by calcineurin/NFAT pathway and T cell receptor (TCR) signalling in primary human T cells.

Conclusion *LINC01871* influences expression of many immune cell genes and growth factors, is IFN γ inducible, and regulated by calcineurin signalling and TCR ligand engagement. Altered *LINC01871* expression may influence the dysregulated T cell inflammatory pathways implicated in SjD.

INTRODUCTION

Sjögren's disease (SjD) is a heterogeneous autoimmune disease that affects ~0.7% of the worldwide population with a female-to-male

WHAT IS ALREADY KNOWN ON THIS TOPIC

- ⇒ Sjögren's disease (SjD) is an understudied heterogeneous autoimmune disease generally characterised as extreme dryness of the eyes and mouth caused by immune-mediated destruction of exocrine glands.
- ⇒ Identifying transcripts that are differentially expressed in SjD subphenotypes and have functional implications in disease pathology (especially undercharacterised long non-coding RNAs (lncRNAs)) provides important new insights into the disease mechanisms that drive SjD and identify potential biomarkers of different subphenotypes.

WHAT THIS STUDY ADDS

- ⇒ Pathway analyses of whole blood transcriptomes from patients with SjD that are either positive or negative for the hallmark autoantibody, anti-Ro (SjD^{Ro+} vs SjD^{Ro-}), and healthy controls suggest these two SjD subphenotypes may have different mechanisms of disease pathology: interferon-mediated disease in SjD^{Ro+} patients and T cell-driven disease in SjD^{Ro-} and SjD^{Ro+} patients.
- ⇒ Identified the lncRNA, *LINC01871*, as differentially expressed in SjD and important for T cell function.

HOW THIS STUDY MIGHT AFFECT RESEARCH, PRACTICE OR POLICY

- ⇒ Study clearly implicates dysregulation of lncRNAs as important mechanisms driving both interferon-mediated and T cell-driven pathology of SjD. As such, it provides critical insights for future studies aimed at identifying biomarkers of SjD for clinical diagnosis. The approaches described herein also provide a roadmap to guide future studies in the discovery and functional characterisation of novel lncRNAs with potential disease implications.

disparity of greater than 9:1.^{1–3} Extreme dry eyes and mouth are hallmark symptoms of SjD caused by chronic inflammation, lymphocytic

infiltration, and subsequent destruction of lacrimal and salivary glands.^{4,5} Many patients with SjD also experience extraglandular complications including pulmonary and kidney dysfunction, neuropathy, debilitating fatigue, arthritis/arthralgia, leucocytoclastic vasculitis and an increased risk of 9-fold to 20-fold of non-Hodgkin's lymphoma.⁵ Although the aetiology and pathological mechanisms remain unknown, it is widely hypothesised that genetic susceptibility likely influences disease onset, progression and heterogeneity in the context of specific, but not yet defined, environmental conditions.^{6,7}

Research classification criteria for SjD require focal lymphocyte sialoadenitis of a biopsied salivary gland and/or circulating autoantibodies to the RNA-binding protein, Ro60/SSA/TROVE-2 and/or the ubiquitin ligase protein, Ro52/TRIM21.^{8–11} Circulating anti-Ro/SSA antibodies are common in SjD (~60%–70%; SjD^{Ro+}) and have been correlated with the upregulation of interferon (IFN)-stimulated genes (ie, IFN signature), increased lymphocyte infiltration of glands, more severe salivary gland involvement, increased prevalence of systemic extraglandular disease and higher risk of developing non-Hodgkin's lymphoma.^{12–14} In contrast, patients with anti-Ro autoantibody-negative SjD (SjD^{Ro-}) lack the IFN signature and have a lower risk of lymphoma, but exhibit increased dryness measures, peripheral nervous system involvement and evidence of T cell-driven pathologies.^{15–17}

Long non-coding RNAs (lncRNAs) are a diverse group of non-protein-coding RNAs greater than 200 nucleotides in length that modulate chromatin remodelling, transcription, and/or post-transcriptional modifications.¹⁸ Transcriptome profiling studies have identified numerous differentially expressed (DE) lncRNAs in autoimmune diseases, including SjD.^{19–25} These and similar studies applied expression correlation analyses with protein-coding RNAs (pcRNAs) to gain insights into the molecular roles of DE lncRNAs, but precise functional mechanisms remain undefined for many of the ~150 000 reported lncRNAs in the NONCODE database (noncode.org). For example, the lncRNA, *negative regulator of the IFN response (NRIR)*, was shown to impair hepatocyte responses to IFN, but was also reportedly upregulated and functioned to positively regulate IFN responses in monocytes from patients with systemic sclerosis.^{26,27} *Nuclear enriched abundant transcript 1 (NEAT1)* positively regulates BAFF-mediated type I IFN activation of B cells in lupus-prone mice, mitogen-activated protein kinase (MAPK) signalling in cells from patients with SjD and inflammation signalling through TLR4.^{22,28,29} *IFNG-AS1 (TMEVPG1 or NeST)* increases Th1 responses in patients with SjD.³⁰ Together, these studies suggest previously uncharacterised DE lncRNAs may yield important mechanistic implications and/or serve as novel biomarkers for SjD and other human diseases.

This study leveraged whole blood RNA-sequencing (RNA-seq) analyses coupled with bioinformatics pathway and co-expression correlation analyses to

identify and differentiate important regulators of the IFN signature-dominant SjD^{Ro+} subphenotype from the T cell-driven pathologies shared by SjD^{Ro+} and SjD^{Ro-}. To this end, we identified four antisense lncRNAs that were overexpressed in the SjD^{Ro+} and co-expressed with IFN-regulated transcripts. *LINC01871*, a previously uncharacterised lncRNA, was found to be overexpressed in both SjD subphenotypes and regulated through both IFN γ and calcineurin/NFAT signalling. Collectively, this study provides insights into the pathways that influence SjD^{Ro+} and SjD^{Ro-} subphenotypes and mechanistic roles of several lncRNAs implicated in immune cell regulation and SjD pathogenesis.

METHODS

See online supplemental materials and methods.

RESULTS

RNA-seq was performed using RNA from globin-depleted whole blood from 50 patients with SjD (27 SjD^{Ro+}, 23 SjD^{Ro-}) and 27 healthy controls (HCs) (online supplemental table 1). After quality control, a total of 37 821 transcripts were analysed for DE (\log_2 fold change (\log_2 FC) ≥ 1 or ≤ -1 ; false discovery rate-adjusted p value (p_{adj}) ≤ 0.05). Four analyses were performed: (1) all SjD cases (SjD^{All}) versus HCs (figure 1A; online supplemental table 2); (2) SjD^{Ro+} versus HCs (figure 1B; online supplemental table 3); (3) SjD^{Ro-} versus HCs (figure 1C; online supplemental table 4); (4) SjD^{Ro+} versus SjD^{Ro-} (online supplemental table 5). The SjD^{All} versus HC, SjD^{Ro+} versus HC and SjD^{Ro-} versus HC analyses identified 4451, 2377, and 6379 DE transcripts, respectively, with nearly equal representation of pcRNAs and lncRNAs (figure 1D–F; online supplemental table 6). The majority of genes were downregulated in all three SjD DE transcript sets relative to HCs (figure 1D–F), indicating repression of gene expression, lack of appropriate upregulation of gene expression or substantially different blood cellular composition in SjD. Differential blood cellularity, mostly involving leucocytopenia, has been described in SjD.^{16,31} Deconvolution was performed using quantiseq³² to assess whether differences in cell type composition between the three datasets may confound interpretations of the RNA-seq results. Significant differences between SjD^{Ro+} or SjD^{Ro-}, and HCs were only observed in macrophages and monocytes (online supplemental figure 1).

Differential expression of pcRNAs

A total of 3097 unique pcRNAs were DE across all three analyses (figure 1E). In the SjD^{All} versus HC and SjD^{Ro+} versus HC analyses, the most upregulated DE pcRNAs were IFN-stimulated genes: *RSAD2*, *IFI44L* and *IFI27* (figure 1A,B; table 1; online supplemental tables 2 and 3).³³ The IFN-stimulated gene, *S100A8*, was upregulated in all three analyses. In contrast, many of the upregulated DE pcRNAs in the SjD^{Ro-} versus HC analysis, including *ATG10* and *UQCRRB*, were involved in protein processing

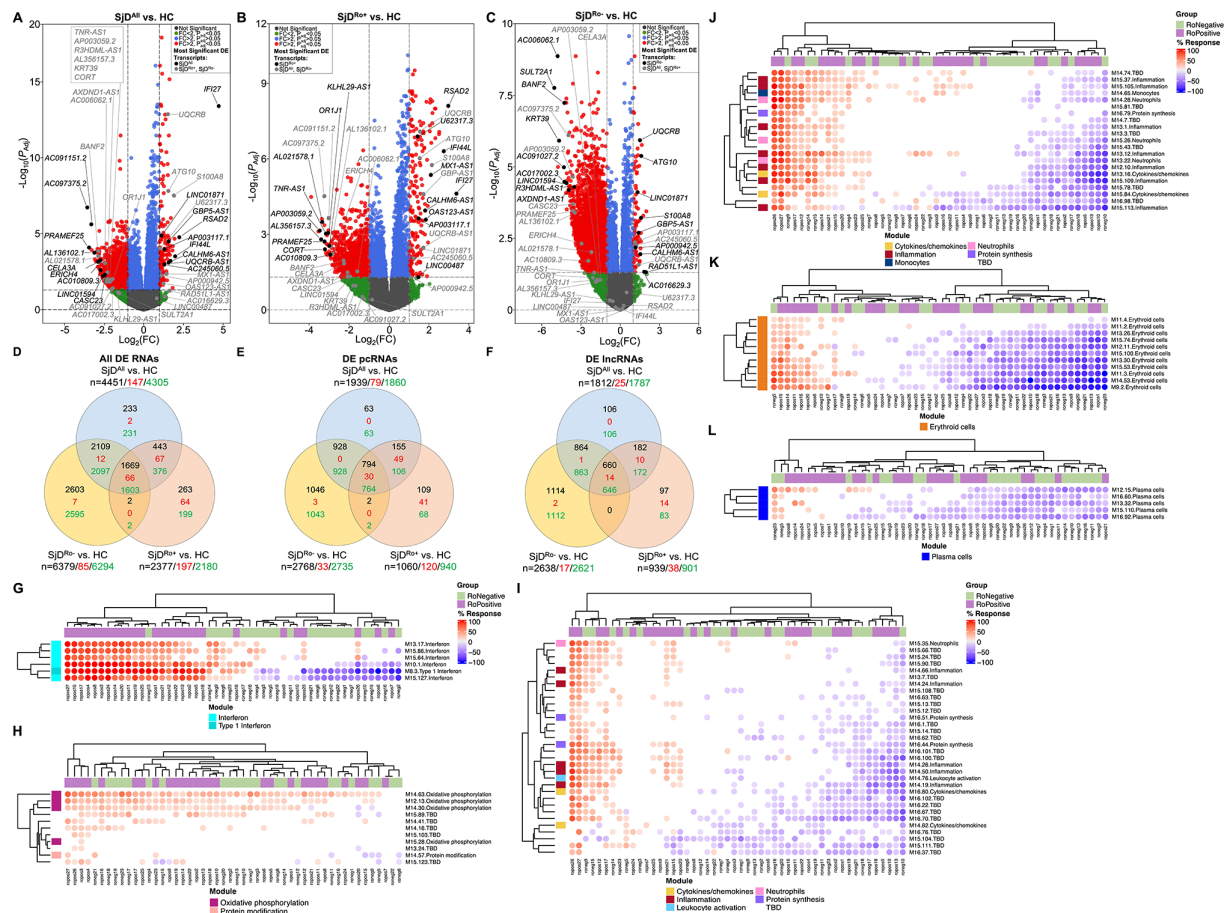


Figure 1 Protein-coding (pc)RNAs and long non-coding (lnc)RNAs are differentially expressed in the whole blood of SjD cases compared with healthy controls (HCs). (A–C) Differentially expressed (DE) transcripts from whole blood RNA-sequencing analysis of (A) all SjD cases (SjD^{All}; n=50), (B) anti-Ro positive SjD cases (SjD^{Ro+}; n=27) or (C) anti-Ro negative SjD cases (SjD^{Ro-}; n=23) compared with HCs (n=27). Y-axis shows the $-\log_{10}$ of the FDR-adjusted p value (p_{adj}); x-axis shows the \log_2 of the fold change (FC). Black dots indicate the top three upregulated and downregulated pcRNAs and lncRNAs in the analysis. Grey dots indicate the top three upregulated and downregulated pcRNAs and lncRNAs in the other analyses. (D–F) Distribution of (D) all DE transcripts, (E) DE pcRNAs or (F) DE lncRNAs across the three analyses. Black text indicates total DE transcripts; red text indicates upregulated transcripts; green text indicates downregulated transcripts. (G–L) Hierarchical clustering of annotated module aggregates across individual SjD^{Ro+} (purple) or SjD^{Ro-} (green) cases. Displayed individual modules had DE of >20% of constitutive transcripts in at least one case. Colour gradient indicates the proportion of DE transcripts ranging from 100% increased (red) to 100% decreased (blue), respective to HCs. FDR, false discovery rate; SjD, Sjögren's disease.

and cell metabolism (figure 1C; table 1; online supplemental table 4).

Many of the most downregulated DE pcRNAs from the SjD^{Ro-} versus HC analysis were also DE in the SjD^{All} versus HC and SjD^{Ro+} versus HC analyses (figure 1A–C; table 1), indicative of common pathways across disease subphenotypes. Further, several of these transcripts, including *CORT*, *PRAMEF25*, *KRT39*, *ERICH4* and *BANF2*, have not been previously reported as DE in SjD.

Pathway analyses of DE transcripts

Gene Set Enrichment Analysis^{34 35} (online supplemental tables 7 and 8), Ingenuity Pathway Analysis (IPA) (online supplemental figure 2A and tables 9–11) and the BloodGen3 module approach previously developed for whole blood lupus studies^{36–38} (figure 1G–L; online supplemental figure 2B–D) were performed using the DE transcript lists to gain insights into the functional

similarities and differences between SjD^{Ro+} and SjD^{Ro-}. Consistent with previous reports, the pathway analyses of the DE transcripts from the SjD^{Ro+} versus HC analysis strongly indicated upregulated type I IFN signalling and other pro-inflammatory pathways (figure 1G,I,J; online supplemental figure 2A–C and table 12). Pathways of significance for the SjD^{Ro-} versus HC analysis included antigen receptor, cytokine and calcium/cAMP signalling (online supplemental figure 2A,B,D and table 12). Examination of all gene module sets from the BloodGen3 approach revealed significant heterogeneity among individual cases and, apart from the type I IFN signature, no clear segregation based on autoantibody status. The majority of subjects with SjD did exhibit increased expression of genes in the oxidative phosphorylation module set (figure 1H). Further, a subset of subjects with SjD, irrespective of autoantibody status, showed

Table 1 RNA-seq analysis of whole blood from SJD cases reveals common and subphenotype-specific differentially expressed pcRNAs and lncRNAs

ENSEMBL ID	Gene symbol	Cytoband	Strand	Chromosome position (hg19)	SJD ^{All}			SJD ^{Ro+}			SJD ^{Ro-}		
					Log ₂ FC	P _{adj}		Log ₂ FC	P _{adj}		Log ₂ FC	P _{adj}	
pcRNA	ENSG00000143546	S100A8	1q21.3	-	chr1:153,362,508-153,363,664	1.91	3.03E-08	2.13	1.75E-06	1.60	6.17E-04		
	ENSG00000137959	IFI44L	1p31.1	+	chr1:78,619,922-78,646,145	1.99	3.09E-04	2.72	4.98E-07	0.02	9.60E-01		
	ENSG00000134321	RSAD2	2p25.2	+	chr2:6,865,806-6,898,239	2.25	1.69E-05	2.95	7.82E-09	0.24	5.79E-01		
	ENSG00000152348	ATG10	5q14.1	+	chr5:81,972,023-82,276,857	1.56	1.52E-08	1.53	9.35E-07	1.60	4.07E-06		
	ENSG00000156467	UQCRB	8q22.1	-	chr8:96,222,947-96,235,546	1.52	1.35E-13	1.54	8.08E-08	1.51	1.14E-06		
	ENSG00000165949	IFI27	14q32.12	+	chr14:94,104,836-94,116,698	4.74	3.90E-14	3.36	2.39E-05	-0.40	3.76E-01		
	ENSG00000241563	CORT	1p36.22	+	chr1:10,450,031-10,451,998	-2.59	8.40E-03	-3.22	3.95E-03	-2.07	5.69E-02		
	ENSG00000229571	PRAMEF25	1p36.21	+	chr1:13,068,677-13,077,884	-3.46	8.10E-05	-3.24	1.95E-03	-3.71	2.87E-04		
	ENSG00000142789	CELA3A	1p36.12	+	chr1:22,001,657-22,012,542	-2.87	5.61E-04	-2.16	1.32E-02	-1.57	7.87E-02		
	ENSG00000136834	OR1J1	9q33.2	-	chr9:122,476,958-122,477,926	-2.35	8.86E-03	-3.06	8.80E-04	-1.77	6.25E-02		
	ENSG00000196859	KRT39	17q21.2	-	chr17:40,958,417-40,966,948	-2.43	1.51E-02	-1.72	9.07E-02	-4.65	1.20E-06		
	ENSG00000204978	ERICH4	19q13.2	+	chr19:41,443,156-41,444,765	-3.02	6.33E-04	-2.97	5.99E-03	-3.06	3.75E-03		
	ENSG00000105398	SULT2A1	19q13.33	-	chr19:47,870,467-47,886,315	-1.66	7.15E-02	-0.79	3.96E-01	-5.02	1.70E-08		
	ENSG00000125888	BANF2	20p12.1	+	chr20:17,693,672-17,735,871	-2.83	1.05E-04	-2.18	7.37E-03	-4.23	5.65E-08		
	ENSG00000237568	GBP5-AS1	1p22.2	+	chr1:89,260,582-89,269,754	1.58	3.20E-05	1.72	2.54E-05	1.42	1.99E-03		
	ENSG00000244158	CALHM6-AS1	6q22.1	-	chr6:116,460,739-116,463,692	1.68	6.12E-04	1.80	1.13E-04	1.54	1.12E-02		
	ENSG00000254224	UQCRB-AS1	8q22.1	+	chr8:96,235,427-96,239,149	1.57	8.02E-04	1.70	3.39E-03	1.41	2.64E-02		
	ENSG00000257452	OAS123-AS1	12q24.13	-	chr12:112,907,628-113,017,751	1.24	1.83E-02	1.83	2.61E-04	0.00	9.98E-01		
	ENSG00000259038	RAD51L1-AS1	14q24.1	-	chr14:68,627,166-68,628,445	1.64	5.64E-02	1.28	NR	1.98	4.80E-02		
	ENSG00000228318	MX1-AS1	21q22.3	-	chr21:42,813,321-42,814,669	1.43	4.44E-03	2.07	4.36E-06	-0.09	8.84E-01		
	ENSG00000260990	TNR-AS1	1q25.1	+	chr1:175,307,218-175,335,459	-2.52	6.47E-03	-3.16	9.46E-04	-1.99	3.62E-02		
Antisense lncRNA	ENSG00000261250	AXDND1-AS1	1q25.2	-	chr1:179,543,201-179,548,922	-2.43	3.35E-03	-1.91	2.86E-02	-3.68	5.32E-05		
	ENSG00000224361	KLHL29-AS1	2p24.1	-	chr2:23,507,043-23,524,344	-1.68	5.51E-02	-3.09	2.49E-03	-0.82	3.93E-01		
	ENSG00000225328	LINC01594	2q12.3	-	chr2:108,167,125-108,217,886	-2.57	4.91E-03	-1.87	4.83E-02	-4.03	2.94E-05		
	ENSG00000255420	CASC23	11p15.4	-	chr11:8,011,278-8,016,520	-2.53	4.46E-03	-1.96	4.61E-02	-3.59	1.97E-04		
	ENSG00000276702	AC010809.3	15q14	-	chr15:33,850,538-33,851,178	-2.80	2.14E-03	-3.08	5.63E-03	-2.52	1.72E-02		
	ENSG00000226812	R3HDM1-AS1	20q13.12	-	chr20:44,347,552-44,355,185	-2.40	8.07E-03	-1.65	9.74E-02	-3.92	6.23E-05		

Continued

Table 1 Continued

ENSEMBL ID	Gene symbol	Cytoband	Strand	Chromosome position (hg19)	SjD ^{All}			SjD ^{Ro+}			SjD ^{Ro-}		
					Log ₂ FC	P _{adj}		Log ₂ FC	P _{adj}		Log ₂ FC	P _{adj}	
lncRNA	ENSG00000235576	LINC01871	2p25.1	+	chr2:7,725,801–7,730,705	1.16	1.78E-05	1.01	3.26E-03	1.33	7.67E-05		
	ENSG00000205837	LINC00487	2p25.2	–	chr2:6,728,177–6,770,311	0.91	1.75E-01	1.49	4.04E-02	–0.31	7.12E-01		
	ENSG00000270077	AP003117.1	8p23.1	+	chr8:97,144,170–97,144,723	1.33	1.02E-04	1.47	2.31E-04	1.15	3.79E-03		
	ENSG00000288528	AP000942.5	11q22.2	–	chr11:102,316,173–102,328,586	1.13	5.14E-03	1.04	4.99E-02	1.25	5.54E-03		
	ENSG00000286104	AC016629.3	19q13.43	+	chr19:58,575,445–58,601,316	1.25	1.33E-01	0.47	NR	1.84	4.71E-02		
	ENSG00000274422	AC245060.5	22q11.22	–	chr22:22,283,928–22,287,220	1.22	6.99E-04	1.22	9.18E-03	1.22	5.39E-03		
	ENSG00000273272	U62317.3	22q13.33	+	chr22:50,541,108–50,543,011	1.03	8.18E-05	1.44	1.67E-07	0.36	1.83E-01		
	ENSG00000285016	AC017002.3	2q13	–	chr2:111,429,324–111,699,033	–2.01	3.03E-02	–1.21	2.16E-01	–4.24	3.27E-05		
	ENSG00000249818	AC097375.2	4q31.3	–	chr4:151,904,932–151,928,710	–3.56	1.87E-07	–3.25	3.00E-05	–3.94	7.68E-07		
	ENSG00000287173	AL356157.3	10q11.21	+	chr10:44,509,708–44,515,421	–2.48	1.26E-02	–3.41	1.56E-03	–1.73	1.01E-01		
	ENSG00000255250	AP003059.2	11q14.2	+	chr11:86,727,355–86,765,467	–2.40	7.75E-03	–3.49	7.12E-04	–1.64	8.11E-02		
	ENSG00000267354	AC091151.2	18q12.3	+	chr18:45,507,202–45,550,183	–3.31	2.36E-06	–3.03	2.24E-04	–3.64	1.93E-05		
	ENSG00000266586	AC091027.2	18q12.3	–	chr18:78,908,629–78,918,993	–2.04	3.21E-02	–1.20	2.28E-01	–4.28	1.01E-05		
	ENSG00000275894	AL021578.1	20q13.12	–	chr20:45,345,115–45,345,823	–2.87	2.91E-04	–3.35	3.18E-04	–2.38	8.14E-03		
	ENSG00000287107	AL136102.1	20q13.13	+	chr20:48,120,269–48,136,844	–3.12	1.75E-04	–2.86	3.48E-03	–3.50	3.28E-04		
	ENSG00000287171	AC006062.1	Xq21.1	+	chrX:8,487,165–8,504,564	–2.43	7.94E-04	–1.61	2.62E-02	–4.76	1.32E-09		

Bold indicates top three upregulated and downregulated pcRNAs and lncRNAs in each analysis.

FDR, false discovery rate; lncRNAs, long non-coding RNAs; Log₂FC, log₂ fold change; P_{adj}, FDR-adjusted p value; pcRNAs, protein-coding RNAs; RNA-seq, RNA sequencing; SjD, Sjögren's disease; SjD^{All}, all SjD cases; SjD^{Ro-}, anti-Ro negative SjD; SjD^{Ro+}, anti-Ro positive SjD.

increased expression of genes in the erythroid module set (figure 1K), perhaps consistent with recent work showing the presence of erythrocyte mitochondria in juvenile systemic lupus erythematosus.³⁹ Collectively, these analyses suggest that the disease pathologies of SjD^{Ro+} involve a prominent IFN signature that is absent from, and less likely to influence disease pathology of, SjD^{Ro-}. However, the heterogeneous stratification of SjD^{Ro+} and SjD^{Ro-} subjects with respect to other gene module sets suggests that multiple mechanisms likely contribute to SjD, with shared mechanisms of cellular metabolism and immune cell dysfunction.

Differential expression of lncRNAs

A total of 660 unique DE lncRNAs were shared across all three whole blood RNA-seq analyses; 97 lncRNAs were unique in the SjD^{Ro+} versus HCs; 1114 lncRNAs were unique in the SjD^{Ro-} versus HCs (figure 1F; online supplemental table 6). The most upregulated DE antisense lncRNAs in the SjD^{Ro+} versus HCs were also upregulated in the SjD^{All} versus HC analysis and are antisense to pcRNAs implicated in type I IFN signalling: *AC004551.1* (antisense spanning the genes *OAS1*, *OAS2* and *OAS3*, therefore, named *OAS123-AS1*; ENSG00000257452) and *AP001610.1* (directly overlapping antisense to *MX1*, therefore, named *MX1-AS1*; ENSG00000228318) (figure 1A,B; table 1; online supplemental tables 2 and 3).³³ The IFN-inducible lncRNA, *NRIR*,^{26 27} was also upregulated in both SjD^{All} versus HCs and SjD^{Ro+} versus HCs (online supplemental tables 2 and 3). Significant upregulation of *NRIR*, *OAS123-AS1* and *MX1-AS1* in SjD^{All} and SjD^{Ro+} was validated by quantitative reverse transcription PCR (RT-qPCR) in an independent cohort of 22 cases with SjD (14 SjD^{Ro+}, 8 SjD^{Ro-}) and 24 HCs (online supplemental figure 3A–C). *AC099063.1* (directly overlapping antisense to the type II IFN responsive gene, *GBP5*; therefore, named *GBP5-AS1*; ENSG00000237568), *CALHM6-AS1* and *UQCRB-AS1* were upregulated DE antisense lncRNAs in all three RNA-seq analyses (figure 1A–C; table 1; online supplemental tables 2–4). *GBP5-AS1* upregulation was validated by RT-qPCR in the independent SjD^{Ro+} and trended in SjD^{All} (online supplemental figure 3E). As observed with the pcRNAs, many of the downregulated antisense lncRNAs were shared among the three analyses, such as *TNR-AS1*, *AXDND1-AS1* and *CASC23* (figure 1A–C; table 1; online supplemental tables 2–4).

Several lncRNAs, including *LINC01871* and *AC245060.5*, were also DE across all three RNA-seq analyses (figure 1A–C; table 1; online supplemental tables 2–4). *LINC01871* upregulation was validated by RT-qPCR in the independent cohort (online supplemental figure 3D). Two DE lncRNAs were uniquely upregulated in the SjD^{Ro+} versus HCs: *LINC00487* and *U62317.3* (figure 1B; table 1; online supplemental table 3). *AC016629.3* was unique to and the most upregulated DE lncRNA in the SjD^{Ro-} versus HCs (figure 1C; table 1; online supplemental table 4). Lastly, the SjD^{Ro-} versus HC analysis exhibited the most downregulated lncRNAs with two

subphenotype-specific transcripts: *AC017002.3* and *AC091027.2* (figure 1C; table 1; online supplemental table 4).

Identification and cell type-specific expression of IFN-related lncRNAs

Pearson's correlation analyses were performed using the highly DE IFN-stimulated pcRNA, *RSAD2* and DE transcript lists from the SjD^{All}, SjD^{Ro+}, SjD^{Ro-}, and HC data-sets to identify *RSAD2*-correlated transcripts ($r > 0.65$ or < -0.6 ; $p < 0.05$) (figure 2A; online supplemental table 13). Numerous IFN-responsive pcRNAs and 16 lncRNAs from the SjD^{Ro+} DE transcript list correlated with *RSAD2* expression, including *MX1-AS1*, *NRIR*, *BISPR*, *OAS123-AS1* and *IRF1-AS1* (figure 2A; online supplemental tables 13 and 14).^{33 40} *CYTOR*, an lncRNA implicated in cancer and autophagy,^{41–43} was also correlated. Five anti-correlated lncRNAs were also identified, including *ZNF793-AS1*, which was DE in SjD^{All} versus HC and SjD^{Ro-} versus HC RNA-seq analyses.

Individual-level expression of transcripts that were both DE in the whole blood RNA-seq analyses and correlated with *RSAD2* expression segregated into IFN high, moderate and low clusters that corresponded with anti-Ro antibody status (figure 2B). Correlated expression of *RSAD2* and IFN-responsive lncRNAs in SjD^{Ro+} subjects further implicated these lncRNAs as potential regulators of IFN responses. Interestingly, Blueprint Epigenome cell type-specific RNA-seq data from 62 cell types revealed lower and more cell type-restricted expression of these lncRNAs, relative to IFN-responsive pcRNAs (online supplemental figure 3G).

Modulation of lncRNAs in response to IFN in vitro

RT-qPCR was performed with RNA from Kasumi-3 (early myeloid) cell lines stimulated with IFN α or IFN γ over a 48-hour time course to determine if the *RSAD2* correlated and DE lncRNAs were modulated by IFN. *NRIR* and the IFN-stimulated pcRNAs, *RSAD2* and *CMPK2* were coordinately upregulated in response to IFN α in Kasumi-3 cells (figure 2C). *OAS123-AS1*, *MX1-AS1* and *GBP5-AS1* were also upregulated, but their induction preceded upregulation of respective pcRNAs, *OAS1*, *MX1* or *GBP5* (figure 2C). IFN γ stimulation also upregulated *NRIR* expression by 6 hours, and *OAS123-AS1*, *MX1-AS1* and *GBP5-AS1* expression by 1.5 hours (figure 2D). Observed kinetics of the IFN response suggests that these antisense lncRNAs may positively regulate the spatially linked pcRNAs.

Identification of dysregulated lncRNAs common across SjD subphenotypes

Identifying and characterising lncRNAs that are common among different SjD subphenotypes may provide new insights into common disease mechanisms. Fourteen of the 660 shared DE lncRNAs identified in the whole blood RNA-seq analyses were overexpressed (figure 1F; online supplemental table 6). Twelve were

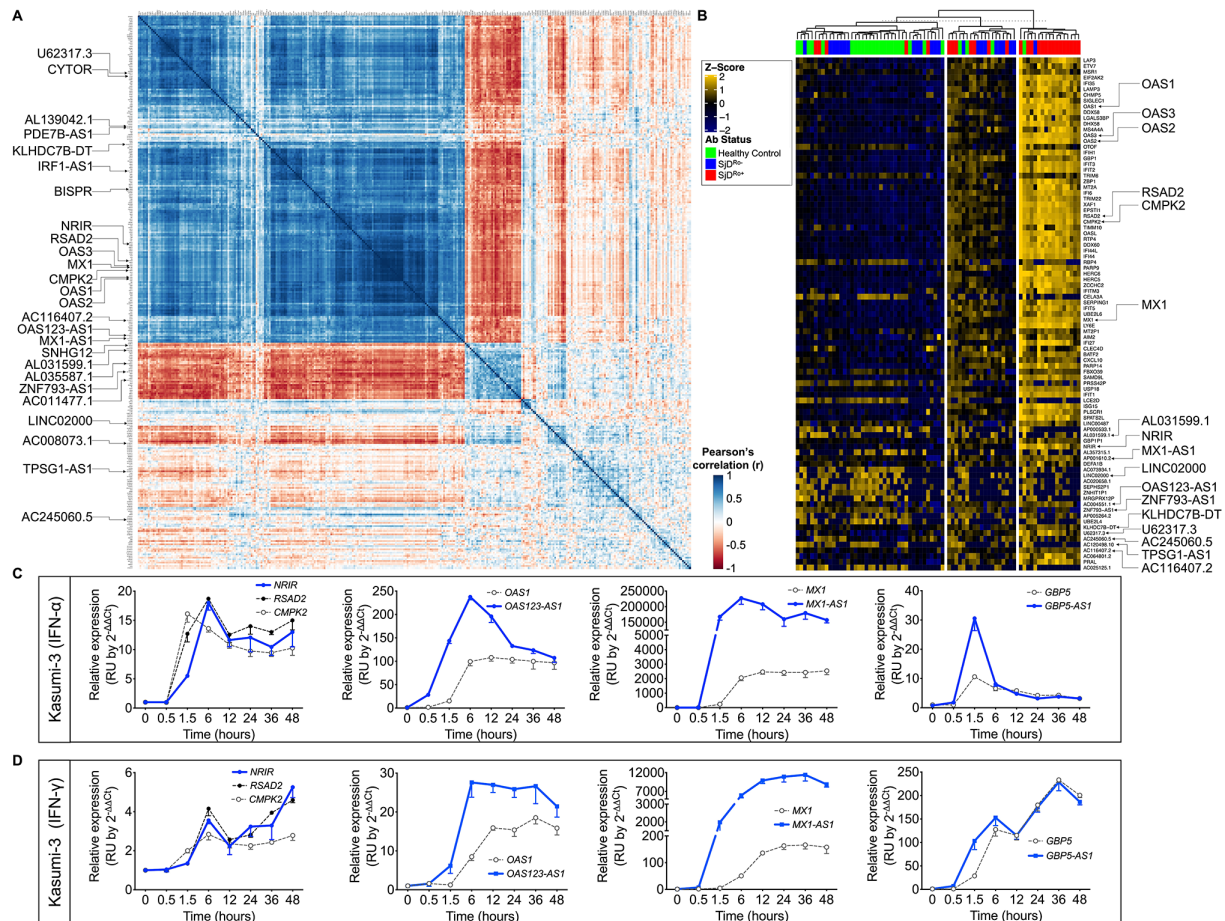


Figure 2 Differentially expressed (DE) interferon (IFN)-responsive lncRNAs were coordinately modulated with associated pcRNAs and correlated with antibody status of SjD cases. (A) *RSAD2* correlation analysis of the normalised RNA-seq data from the SjD^{Ro+}-only primary expression matrix ($r \geq 0.65$ or ≤ -0.6 ; $p < 0.05$). Type I IFN-responsive pcRNAs and other pcRNAs and lncRNAs of interest are indicated. (B) Individual-level expression of transcripts that were correlated with *RSAD2* expression and DE in any of the whole blood RNA-seq analyses. Z-scores were computed using scale function in R, after calculating fold change of expression relative to healthy controls. K-means clustering ($K=3$) segregated cases by high, moderate and low IFN status, and are shown relative to patient antibody status. pcRNAs and lncRNAs of interest are indicated. (C,D) RT-qPCR analysis of indicated DE transcripts in Kasumi-3 cells stimulated with (C) universal type I IFN (IFN α ; 150 U/mL) or (D) IFN γ (2800 U/mL) from 0 to 48 hours. Target lncRNAs are shown in blue and respective pcRNAs in grey ($n > 3$). lncRNAs, long non-coding RNAs; pcRNAs, protein-coding RNAs; RNA-seq, RNA sequencing; RT-qPCR, quantitative reverse transcription PCR; SjD, Sjögren's disease; SjD^{Ro+}, anti-Ro positive SjD; SjD^{Ro-}, anti-Ro negative SjD.

antisense or sense-intronic transcripts, including *GBP5-AS1*, *CALHM6-AS1*, *LARP7-AS1* (*miR302CHG*) and *UQCRB-AS1*, and two were long intergenic non-coding RNAs, *LINC01871* and *AC245060.5* (table 1; online supplemental tables 2–4). Minimal information was available on *AC245060.5*, but *LINC01871* was reported as DE in multiple cancer studies, autoinflammatory diseases, T cells and monocytes (data derived from Gene Expression Atlas; online supplemental figure 3F),^{44–49} making it a transcript of interest in human disease.

LINC01871, located at the 2p25.1 locus, has low exonic conservation among vertebrates, multiple isoforms found only in humans and low protein-coding potential (online supplemental note).⁵⁰ The surrounding genomic interval has an enrichment of H3K27ac epigenetic marks, DNaseI hypersensitivity and transcription factor binding indicative of active transcription. Consistently, *LINC01871* is

strongly expressed in mature CD4⁺ and CD8⁺ T cells, class switched memory B cells, plasma cells, natural killer (NK) cells and haematopoietic progenitor cells (data derived from Blueprint Epigenome RNA-seq data; online supplemental figure 3H). Conversely, regulatory T cells and most B cells have low basal expression of *LINC01871* and early lineage T cells (CD3[–] thymocytes) do not express *LINC01871*. Analysis of publicly available single-cell RNA-seq data of minor salivary gland tissue revealed that *LINC01871* is most highly expressed in tissue-resident T cell subtypes and NK cells (online supplemental figure 3I,J).^{51–52} These data confirm the expression of this lncRNA in immune cells present in SjD-affected tissues, indicating potential relevance to disease pathology.

Interrogation of publicly available GeneFriends,⁵³ FuncPred⁵⁴ and lncRNA2function⁵⁵ databases found that *LINC01871* expression was correlated with pcRNAs

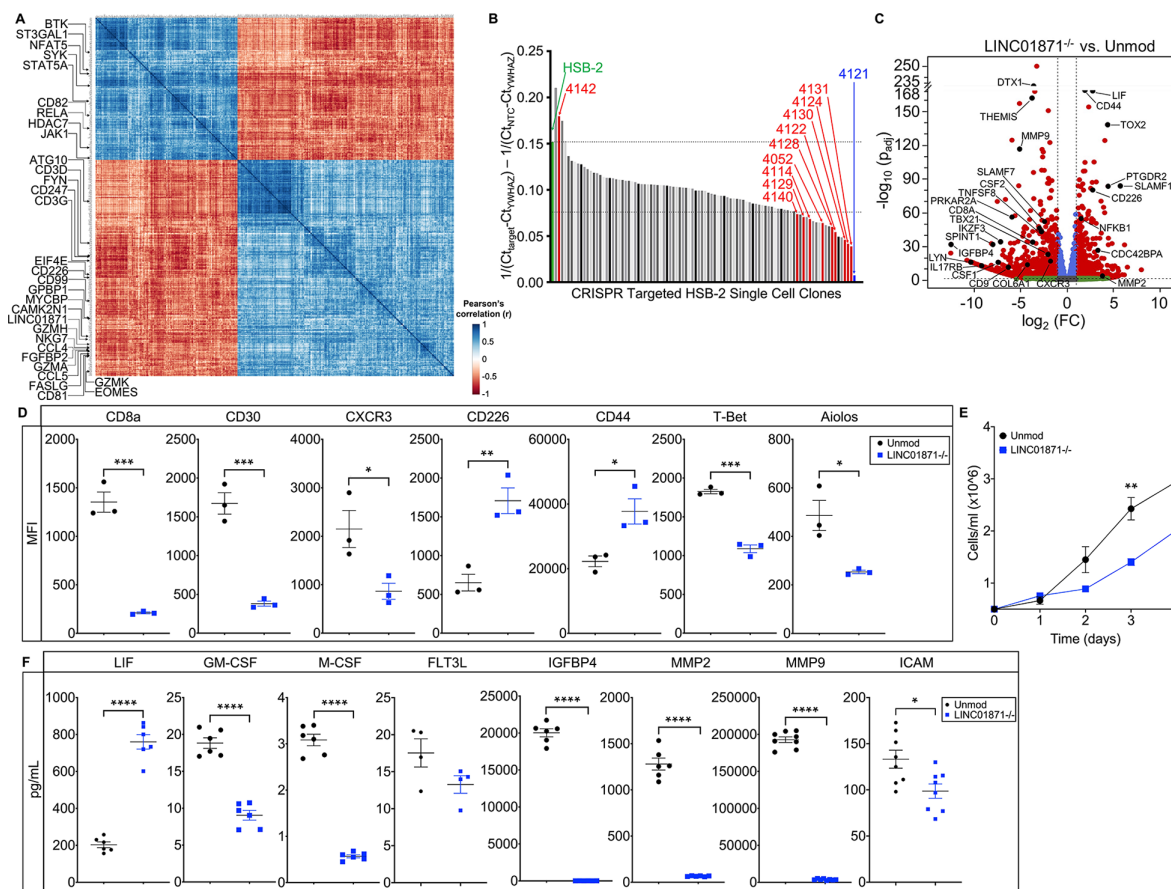


Figure 3 Loss of *LINC01871* disrupts basal expression of genes involved in immune cell regulation. (A) *LINC01871* correlation analysis of normalised RNA-seq data from the SjD^{Ro-} only primary expression matrix ($r \geq 0.7$ or ≤ -0.6 ; $p < 0.05$). Transcripts implicated in immune function or SjD pathology are indicated. (B) Quantitative PCR screen of *LINC01871* expression in HSB-2 single-cell clones after CRISPR-targeted deletion of *LINC01871*. (C) Differentially expressed transcripts from the RNA-seq analysis of HSB-2 clone 4121 (hereafter *LINC01871*^{-/-}) relative to HSB-2 parental cell line. Y-axis shows the $-\log_{10}$ of the FDR-adjusted p value (p_{adj}); x-axis shows the \log_2 of the fold change (FC). Black dots indicate independently replicated transcripts of interest. (D) Levels of indicated surface or intracellular proteins, reported as mean fluorescence intensity (MFI), in HSB-2 parental cells (black) and *LINC01871*^{-/-} cells (blue); $n=3$; unpaired t-test where $*p < 0.05$, $**p < 0.01$ or $***p < 0.001$. (E) Growth curve analysis of HSB-2 parental cells (black) and *LINC01871*^{-/-} cells (blue) from 0 to 96 hours; $n=3$; unpaired t-test where $*p < 0.05$ or $**p < 0.01$. (F) Concentration of indicated secreted protein in supernatant collected from HSB-2 parental cells (black) and *LINC01871*^{-/-} cells (blue) at 96 hours; $n>6$; unpaired t-test where $*p < 0.05$ or $****p < 0.0001$. FDR, false discovery rate; RNA-seq, RNA sequencing; SjD, Sjögren's disease; SjD^{Ro-}, anti-Ro negative SjD.

implicated in IFN γ stimulation, T cell activation and differentiation, myeloid cells and dendritic cells (online supplemental tables 15–17). In this SjD cohort, *LINC01871* expression was highest in the SjD^{Ro-} subjects (online supplemental figure 3D).

Pearson's correlation analyses were performed using the DE transcript list from the SjD^{Ro-} versus HC analysis to identify transcripts of specific interest to immune cell activation and regulation correlated with *LINC01871* ($r > 0.7$ or < -0.6 ; $p < 0.05$) (figure 3A; online supplemental table 18). Of specific interest, *CAMK2N1* and *NFAT5* are both directly regulated by calcium signalling,⁵⁶ while *EOMES*, *GZMA* and *FGFBP2* are important for cytotoxic immune responses.³³ Consistently, Gene Ontology (GO) Enrichment Analysis,⁵⁷ performed using the *LINC01871*-correlated DE transcripts list, identified GO terms implicating *LINC01871* in immune system function, leucocyte

and lymphocyte activation, and IFN γ production (online supplemental table 19).

Functional analysis of *LINC01871* by RNA-seq analysis of a clonal CRISPR-targeted *LINC01871* deletion

To explore the function of *LINC01871*, CRISPR-guided RNAs were designed to target the *LINC01871* interval in early T cell lineage HSB-2 cell line, which constitutively express *LINC01871* (figure 3B; online supplemental figure 4A and table 20). Clones of the CRISPR-edited HSB-2 cells were screened for *LINC01871* deletion by RT-qPCR, identifying clone 4121 (hereafter referred to as *LINC01871*^{-/-}) as having complete deletion of the targeted *LINC01871* interval confirmed by Nanopore sequencing (figure 3B; online supplemental figure 4B,C and tables 21 and 22).

RNA-seq analysis of *LINC01871*^{-/-} cells relative to unedited HSB-2 parental cells identified 1166 DE transcripts ($\log_2\text{FC} \geq 1$ or ≤ -1 ; $p_{\text{adj}} \leq 0.05$) (figure 3C; table 2; online supplemental table 23). Loss of *LINC01871* disproportionately decreased the expression of pcRNAs (~3:1); perturbations of ncRNAs were more equally distributed (online supplemental table 24). Notably, several transcripts that were positively correlated with *LINC01871* in the whole blood RNA-seq analyses, such as *CCL5* ($\log_2\text{FC} = -3.46$; $p_{\text{adj}} = 9.82\text{e-}12$) and *FGFBP2* ($\log_2\text{FC} = -3.54$; $p_{\text{adj}} = 1.62\text{e-}17$) (online supplemental tables 2–4,18), were significantly downregulated in *LINC01871*^{-/-} cells (figure 3C; table 2; online supplemental table 23). In addition, loss of *LINC01871* completely abrogated the expression of 127 transcripts, including adaptive immunity genes *TNFSF8* (CD30L), *IL22*, *CD27* and *LYN* (table 2; online supplemental table 23).

Consistent with the whole blood SjD RNA-seq correlation analyses and database functional predictions, GO Enrichment Analysis and IPA of the *LINC01871*^{-/-} DE transcripts identified several pathways involved in immune cell development and signalling, including Th1/Th2 cells, T/B cell signalling and leucocyte extravasation, as well as pathways impacting growth and cell survival (table 2; online supplemental figure 4D–J and tables 19 and 25). Predictions of such wide-ranging impacts suggest that *LINC01871* may act as a broad regulator impacting many immune cell pathways.

Validation of transcript expression and characterisation of growth and protein expression in *LINC01871*^{-/-} cells

RT-qPCR was used to validate the DE of 32 transcripts using independent biological replicates of the *LINC01871*^{-/-} cells compared with unmodified HSB-2 cells. Twenty-nine of the selected genes, including *LINC01871*, exhibited significant changes in gene expression (online supplemental figure 5A–D). Two of the remaining three transcripts trended but did not reach significance: *SLAMF7* and *CSF2* (online supplemental figure 5A and D). *LINC01871*^{-/-} cells also displayed significant and stable changes in the expression of five cell surface proteins (CD8a, CD30, CXCR3, CD226 and CD44) and two intracellular transcription factors (T-bet and Aiolos) important in immune cell function (figure 3D; online supplemental figure 5E–K).^{58 59}

Given the observed DE of transcripts involved in cell proliferation, the growth characteristics of the *LINC01871*^{-/-} cells were also examined. While early growth rates of the *LINC01871*^{-/-} cells appeared comparable with the unmodified HSB-2 cells, the growth rate of *LINC01871*^{-/-} cells slowed with time in culture, suggesting that deletion of *LINC01871* may perturb the secretion of growth factors (figure 3E). Consistently, several key secreted proteins/growth factors, including *CSF1* (M-CSF protein) and *CSF2* (GM-CSF protein), were reduced in the supernatants from the *LINC01871*^{-/-} cells compared with the unmodified HSB-2 cells (figure 3F; online supplemental figure 5D). IGFBP4, a secreted

inhibitor of insulin-like growth factor (IGF) with functions in inflammation and growth regulation,⁶⁰ was also depleted. *FLT3L*, which encodes the protein Flt3-L, a critical dendritic cell cytokine that acts in concert with GM-CSF,^{61 62} trended toward a decrease. Interestingly, the pleiotropic cytokine, *LIF* (leukaemia inhibitory factor), was markedly increased, implicating *LINC01871* as a negative regulator of this gene.

Perturbations in the concentrations of several proteins commonly associated with cell–cell interactions and extravasation were also observed. The extracellular adhesion and endoprotease proteins, MMP2 and MMP9,^{63 64} were both significantly reduced in the media of *LINC01871*^{-/-} cells (figure 3F). However, *MMP2* gene expression was increased in the RNA-seq analysis ($\log_2\text{FC} = 3.85$; online supplemental figure 5D), suggesting some dysregulation of MMP2 protein secretion in the *LINC01871*^{-/-} cells. Changes in the expression of *MMP9* could not be replicated by qPCR, perhaps due to differential isoform detection by qPCR. Soluble ICAM, a marker of vascular inflammation,⁶⁵ was also significantly decreased. Overall, deletion of *LINC01871* in a T cell line caused widespread disruption of diverse cellular proteins, including the disruption of basal level expression of many proteins involved in adaptive immune responses, cell migration, adhesion and extravasation.

LINC01871 expression is modulated by IFN γ in a myeloid cell line

LINC01871 correlation analyses, bioinformatics data and CRISPR-edited *LINC01871*^{-/-} in vitro studies suggested that *LINC01871* may play a role in both IFN and immune cell activation pathways, specifically IFN γ signalling. Because HSB-2 cells do not respond to in vitro IFN γ stimulations and *LINC01871* was reported to be overexpressed in acute myeloid leukaemia,⁴⁶ we used Kasumi-3 cells to determine if *LINC01871* expression is regulated by universal type I IFN (IFN α) and/or IFN γ (type II IFN) stimulations. IFN α stimulation did not induce *LINC01871* in Kasumi-3 cells, but robustly induced the expression of IFN-inducible genes, *TBX21* and *IRF7* (figure 4A,B). In contrast, IFN γ upregulated *LINC01871* expression (peak 12-fold induction), as well as the IFN-stimulated genes, *TBX21*, *IL6R*, *IDO2* and *IRF7* (figure 4C,D), thus demonstrating that *LINC01871* expression in myeloid cells is regulated by type II IFN signalling.

LINC01871 expression is regulated by calcineurin/NFAT signalling

To further test the hypothesis that *LINC01871* may play a role in adaptive immune cell activation, specifically T cell activation, *LINC01871*^{-/-} and parental HSB-2 cells were stimulated with phorbol 12-myristate 13-acetate and ionomycin (PMA/I) and the modulation of *LINC01871*, *IL2*, *IFNG*, *CD8A*, *TBX21* and *CSF1* expression was assessed by RT-qPCR. PMA/I was used to mimic T cell receptor (TCR) signalling because HSB-2 cells lack a functional TCR.^{66 67} PMA/I increases intracellular calcium without

Table 2 CRISPR deletion of *LINC01871* alters the expression of several immune regulatory transcripts

	Gene (protein) symbol	Log ₂ FC	P _{adj}	1	2	3	4	5	6	7	8	9	10	11	12
Immune regulation and function	LYN	-10.392	1.24E-16								Blue		Purple	Red	
	CD109	-9.707	2.05E-14					Green				Dark Blue			
	TNFSF15	-9.598	4.64E-14	Red					Green						
	IL17RB	-9.324	1.87E-13		Red								Purple		
	IL22	-8.080	1.70E-09								Blue	Dark Blue	Purple	Red	
	IKZF3 (Aiolos)	-7.158	1.62E-34	Red		Yellow		Green			Blue				
	PDCD1 (PD-1)	-6.243	1.07E-06						Green	Cyan	Blue	Dark Blue	Purple	Red	
	CD27	-6.128	1.20E-04	Red	Red	Yellow	Orange				Blue	Dark Blue	Purple		
	TNFSF8 (CD30L)	-5.727	8.29E-04						Green				Purple		
	LAG3	-4.684	1.04E-14				Orange			Cyan		Dark Blue			
	CD79A	-4.091	2.66E-08						Green		Blue		Purple		
	EGR2	-3.925	5.74E-62				Orange				Blue		Purple		
	THEMIS	-3.770	5.85E-162	Red				Green							
	CD8A	-3.683	4.03E-34	Red			Orange					Dark Blue	Purple		
	CCL5	-3.456	9.82E-12	Red	Red	Yellow	Orange					Dark Blue	Purple	Red	
	FOXP3	-3.064	1.78E-03					Green	Green	Cyan	Blue				
	IRF5	-2.959	1.19E-10					Green	Green		Blue		Purple	Red	
	TNFRSF8 (CD30)	-2.537	3.47E-44					Green	Green				Purple		
	SLAMF7	-2.398	1.90E-52				Orange				Blue				
	TBX21 (T-Bet)	-2.066	2.45E-23	Red	Red			Green	Green						
	IL6ST	-1.820	2.16E-21					Green	Green		Blue	Dark Blue	Purple	Red	
	CXCR3	-1.728	2.68E-18	Red				Green	Green			Dark Blue	Purple	Red	
	TNFRSF4	1.910	1.37E-09		Red			Green	Green			Dark Blue			
	CD44	1.924	1.68E-174	Red							Blue	Dark Blue	Purple	Red	
	TGFB3	2.069	8.65E-16		Red		Orange					Dark Blue	Purple	Red	
	CD226	2.750	1.96E-80	Red			Orange		Green			Dark Blue	Purple		
	SELL (CD62L)	3.061	1.82E-15	Red						Cyan	Blue	Dark Blue	Purple	Red	
	HAVCR2	3.139	1.95E-87	Red			Orange		Green	Cyan		Dark Blue		Red	
	CCR4	3.816	4.47E-08	Red	Red								Purple		
	KLF2	4.043	4.13E-60	Red					Green					Red	
	TOX2	4.353	2.58E-138	Red				Green							
	CCR2	4.893	9.69E-13	Red					Green			Dark Blue	Purple	Red	
	TIGIT	5.531	2.31E-08	Red				Green		Cyan					
	SLAMF1 (CD150)	5.726	4.86E-84								Blue	Dark Blue	Purple	Red	
Growth signalling/ cancer regulation	IGFBP4	-12.476	3.61E-32								Blue		Purple	Red	
	CSF1 (M-CSF)	-7.356	5.58E-16								Blue	Dark Blue	Purple	Red	
	PTPN13	-5.998	4.19E-05					Green				Dark Blue	Purple	Red	
	TLR2	-5.914	7.51E-05						Green		Blue	Dark Blue		Red	
	PRKAR2A	-5.899	2.08E-56									Dark Blue	Purple	Red	
	MMP9	-5.096	9.32E-117								Blue	Dark Blue			
	FSCN1	-5.003	0.00E+00					Green						Red	Cyan
	PDGFD	-4.730	7.54E-03				Orange					Dark Blue			
	GZMB	-4.688	2.93E-02				Orange				Blue			Red	
	P2RX7	-4.190	1.17E-08					Green	Green			Dark Blue	Purple	Red	
	ADCY6	-4.163	5.86E-21									Dark Blue	Purple	Red	
	FGFBP2	-3.543	1.62E-17				Orange					Dark Blue			
	CSF2 (GM-CSF)	-3.052	6.62E-48				Orange		Green	Cyan	Blue	Dark Blue	Purple	Red	
	TNF	-1.631	1.22E-88				Orange		Green		Blue	Dark Blue		Red	
	CD28	1.614	7.59E-30	Red	Red				Green		Blue	Dark Blue	Purple		

Continued

Table 2 Continued

Gene (protein) symbol	Log ₂ FC	P _{adj}	1	2	3	4	5	6	7	8	9	10	11	12
XCL1	1.636	1.92E-09												
TLR1	1.674	4.87E-02												
LIF	2.613	2.15E-171												
ITGA6	3.072	0.00E+00												
HPGD	3.517	1.88E-37												
MMP2	3.851	1.17E-02												
ITGAM	4.198	4.15E-03												
IL7R	4.393	1.41E-02												
PTGDR2	4.399	1.27E-83												
ALOX15B	5.189	5.42E-03												
PTGDR	5.460	1.02E-10												
HPGDS	7.992	1.99E-09												

Genes in bold denote correlation with *LINC01871* in SjD^{Ro-} (online supplemental table 18).
 1. Th1 pathway; 2. Th2 pathway; 3. Th1/Th2 activation; 4. CD8 cytotoxic/cytokines; 5. Treg/Th17; 6. T cell development; 7. T cell exhaustion; 8. associated with autoimmune disease;
 9. molecular mechanisms of cancer; 10. proliferation/crosstalk; 11. extravasation/migration; 12. prostaglandin pathway.
 FDR, false discovery rate; log₂FC, log₂ fold change; P_{adj}, FDR-adjusted p value; SjD, Sjögren's disease; SjD^{Ro-}, anti-Ro negative SjD; Treg, regulatory T cell.

surface receptor engagement, leading to activation of the calmodulin-dependent protease, calcineurin, as well as protein kinase C (PKC), and subsequent downstream transcriptional activation through multiple pathways including NFAT and NFκB.^{66 67} *LINC01871* expression was undetectable in *LINC01871*^{-/-} cells. Parental HSB-2 cells showed marked *LINC01871* downregulation after 6 hours of PMA/I stimulation that returned to near basal levels after ~36 hours (figure 4E). Targets of calcineurin/NFAT signalling, *IL2* and *IFNG*, exhibited robust inductions in parental HSB-2 cells, indicating normal stimulation (figure 4F,G). *LINC01871*^{-/-} cells showed an increased induction of *IL2* expression, but an impaired induction of *IFNG* in response to PMA/I (figure 4F,G). This was not due to differences in basal expression as neither transcript was expressed in unstimulated cells. *CD8A* expression increased modestly in HSB-2 cells after PMA/I stimulation but decreased in *LINC01871*^{-/-} cells (figure 4H). *TBX21* and *CSF1* both exhibited a higher magnitude of induction in *LINC01871*^{-/-} cells compared with HSB-2 cells (figure 4I,J); however, this was likely due to lower basal expression in *LINC01871*^{-/-} cells, resulting in a larger fold change induction.

Flow cytometry was used to determine if loss of *LINC01871* also modulated PMA/I-mediated expression of cell surface proteins. PMA/I stimulation induced CD8A surface expression in parental HSB-2 cells but did not change CD8A expression in the *LINC01871*^{-/-} cells, likely because of abrogated CD8A expression in *LINC01871*^{-/-} cells (online supplemental figure 5L). Loss of *LINC01871* prevented the PMA/I-mediated downregulation of CD30 and CXCR3 surface expression observed in parental HSB-2 cells (online supplemental figure 5M,N). In contrast, PMA/I upregulated the surface expression of CD226 and CD44 in both parental HSB-2 and *LINC01871*^{-/-} cells (online supplemental figure 5O,P).

To determine if the PMA/I-mediated downregulation of *LINC01871* resulted from calcineurin activation, parental HSB-2 cells were treated with the calcineurin-specific inhibitor, FK506 (tacrolimus),⁶⁸ or rapamycin (mTOR signalling inhibitor in the same molecular class as FK506), prior to PMA/I stimulation. Treatment with FK506 alone did not alter *LINC01871* expression, but presence of FK506 during PMA/I stimulation restored *LINC01871* expression nearly to that of unstimulated cells (figure 4K). Rapamycin had no effect on *LINC01871* expression. In addition, FK506 mitigated PMA/I-mediated induction of NFAT-regulated genes, *IL2* and *IFNG* (figure 4L,M), and partially inhibited induction of two genes modulated in *LINC01871*^{-/-} cells, *TBX21* and *CSF1* (figure 4O,P). Rapamycin also partially impaired PMA/I-mediated induction of *IL2*, but had no effect on *TBX21* and *CSF1*. Both FK506 and rapamycin caused significant impairment of *IFNG* expression (figure 4M), indicating this pathway is regulated by both calcineurin and mTOR signalling. Neither inhibitor affected PMA/I-induced expression of CD8A (figure 4N). Overall, these data show that *LINC01871* expression is downregulated in T cells in response to direct calcineurin/NFAT activation.

Human T cells regulate *LINC01871* expression by calcineurin/NFAT signalling

To further examine the role of *LINC01871* in T cells, the expression of *LINC01871* and other transcripts of interest were measured in human T cells purified from healthy donor PBMCs treated with FK506 or DMSO and stimulated with PMA/I using RT-qPCR. As in the HSB-2 cells, PMA/I repressed *LINC01871* expression in purified human T cells, beginning at 1.5 hours, and maintained repression 12 hours post-treatment (figure 5A), an effect that was abrogated by FK506. PMA/I-treated human donor T cells also exhibited similar transcriptional

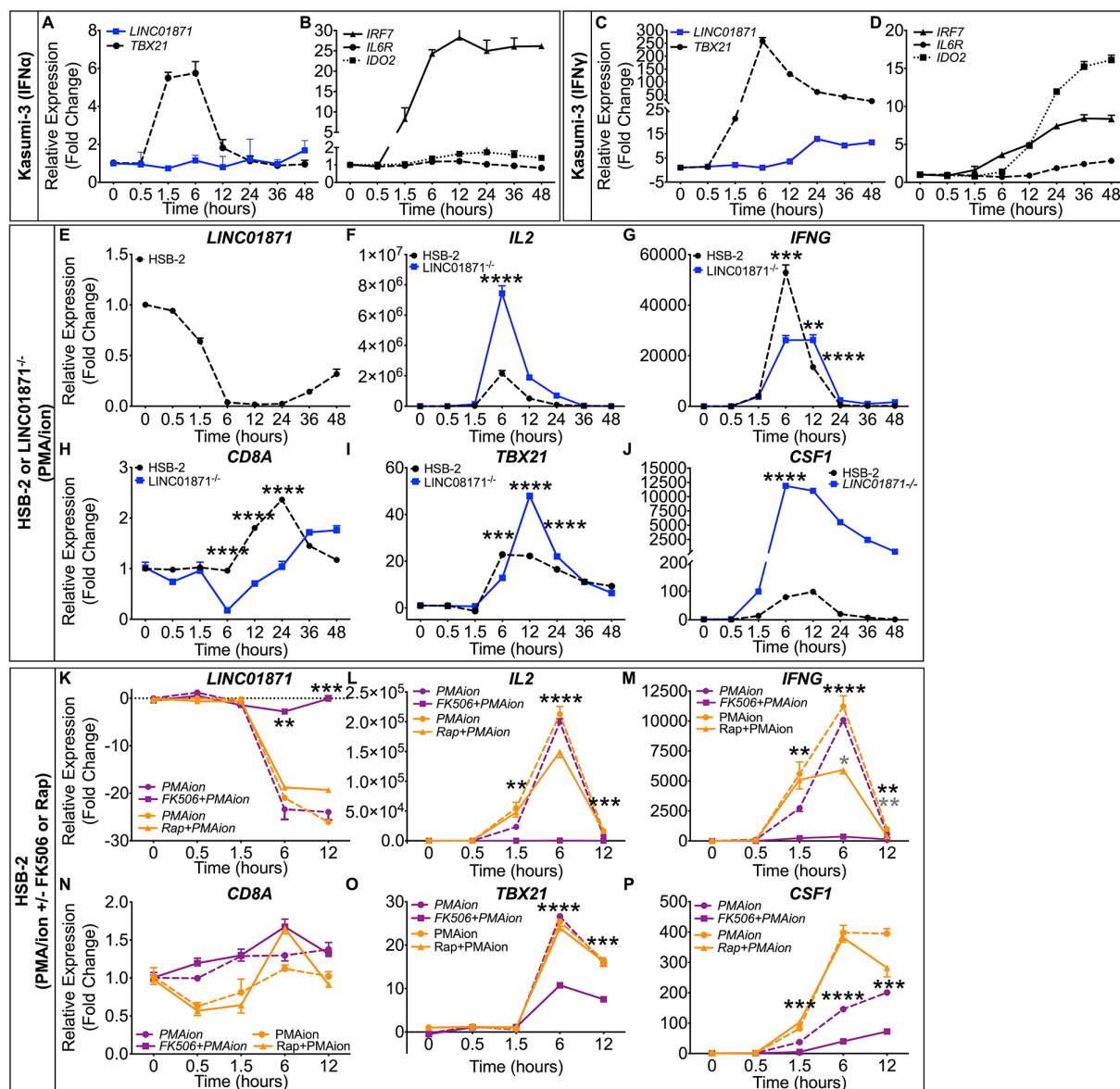


Figure 4 *LINC01871* is modulated by interferon (IFN) γ and calcineurin/NFAT signalling in immune cell lines. (A–D) RT-qPCR analysis of *LINC01871* (A and C), *TBX21* (A and C), *IRF7* (B and D), *IL6R* (B and D) and *IDO2* (B and D) in Kasumi-3 cells stimulated with either (A,B) universal type I IFN (IFN α ; 150 U/mL) or (C,D) IFN γ (2800 U/mL) from 0 to 48 hours. Expression is reported as fold change relative to unstimulated controls. (E–J) RT-qPCR analysis of *LINC01871* (E), *IL2* (F), *IFNG* (G), *CD8A* (H), *TBX21* (I) and *CSF1* (J) in *LINC01871*^{-/-} cells (blue) and HSB-2 parental (black) cells stimulated from 0 to 48 hours with PMA/I (100 ng/mL; 1000 ng/mL). Expression is reported as fold change relative to unstimulated controls; n=3; unpaired t-test where **p<0.01, ***p<0.001 or ****p<0.0001. (K–P) RT-qPCR analysis of *LINC01871* (K), *IL2* (L), *IFNG* (M), *CD8A* (N), *TBX21* (O) and *CSF1* (P) in HSB-2 parental cells stimulated from 0 to 12 hours with PMA/I (100 ng/mL; 1000 ng/mL) with and without FK506 (2.5 μ M; purple) or rapamycin (Rap; 2.5 μ M; orange). Expression is reported as fold change relative to unstimulated inhibitor controls; n=3; paired t-test where *p<0.05, **p<0.01, ***p<0.001 or ****p<0.0001. Black * indicates FK506 with PMA/I relative to PMA/I alone and grey * indicates Rap with PMA/I relative to PMA/I alone. PMA/I, phorbol 12-myristate 13-acetate and ionomycin; RT-qPCR, quantitative reverse transcription PCR.

regulation of *IL2*, *IFNG*, *CD8A* and *TBX21*, relative to HSB-2 cells (figure 5B–E).

Engagement of the TCR triggers a complex signalling cascade, including the activation of multiple signalling effectors including MAPKs, PKC and calcineurin.⁶⁶ In contrast to our findings using PMA/I, *LINC01871* expression was previously shown to increase in response to anti-CD3/CD28-activated TCR signalling in human T cells (online supplemental figure 3F; unpublished data).⁶⁹

To address this, *LINC01871* expression was assessed in purified human T cells stimulated with anti-CD3/CD28 antibodies. TCR signalling decreased *LINC01871* expression at early time points (figure 5F), although to a lower, but significant, degree compared with PMA/I stimulation. However, *LINC01871* expression increased twofold to threefold with longer anti-CD3/CD28 stimulation (48 hours), perhaps mimicking chronic TCR stimulation in vitro. FK506 treatment of anti-CD3/CD28-stimulated

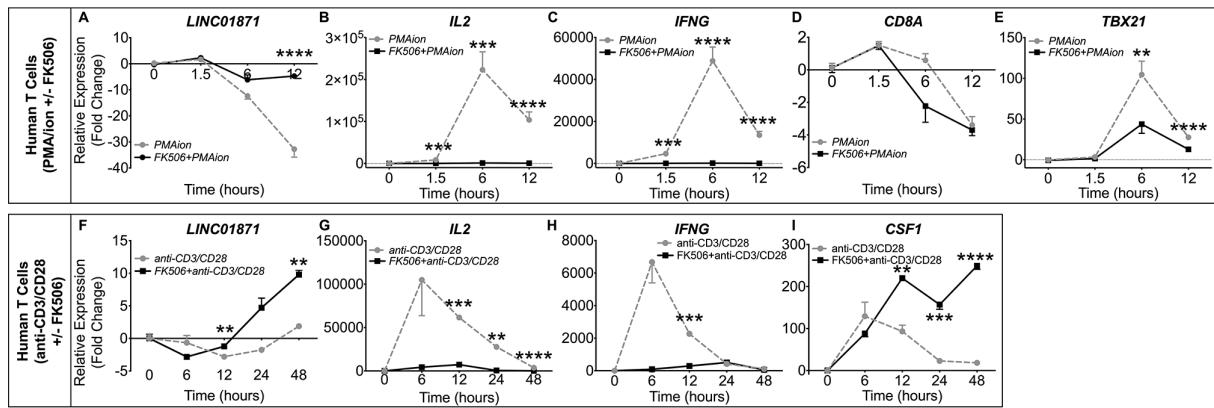


Figure 5 *LINC01871* is modulated by IFN γ and calcineurin/NFAT signalling in purified human T cells. (A–E) Negatively selected human donor T cells were treated with DMSO or 2.5 μ M FK506 overnight, then stimulated with PMA/I (100 ng/mL; 1000 ng/mL) for 12 hours. Expression of *LINC01871* (A), *IL2* (B), *IFNG* (C), *CD8A* (D) and *TBX21* (E) was assessed by RT-qPCR and reported as fold change relative to DMSO-treated cells; n=4; paired t-test where *p<0.05, **p<0.01, ***p<0.001 or ****p<0.0001. Grey lines indicate PMA/I. Black lines indicate PMA/I with FK506 treatment. (F–I) Negatively selected human donor T cells were treated with either DMSO or 2.5 μ M FK506 overnight, then stimulated with anti-CD3/CD28 for 48 hours. Expression of *LINC01871* (F), *IL2* (G), *IFNG* (H) and *CSF1* (I) was assessed by RT-qPCR and reported as fold change relative to DMSO-treated cells at each time point; n=4; paired t-test where *p<0.05, **p<0.01, ***p<0.001 or ****p<0.0001. Grey lines indicate anti-CD3/CD28. Black lines indicate anti-CD3/CD28 with FK506 treatment. IFN, interferon; PMA/I, phorbol 12-myristate 13-acetate and ionomycin; RT-qPCR, quantitative reverse transcription PCR.

T cells shifted the kinetics of downregulation of *LINC01871* at early time points, but augmented expression at late time points. As expected, anti-CD3/CD28 stimulation induced expression of *IL2* and *IFNG*, which was nearly abrogated by FK506 treatment (figure 5G,H). *CSF1* expression also exhibited increased expression with prolonged TCR stimulation in the presence of FK506 treatment (figure 5I). In conclusion, *LINC01871* expression is modulated in human T cells in response to calcineurin/NFAT and TCR signalling. The overexpression of *LINC01871* in SjD, coupled with the alterations observed in *LINC01871*^{-/-} cells and its regulation by both IFN γ and calcineurin/TCR signalling, suggest that this lncRNA plays a role in immune cells that may be relevant to autoimmune disease.

DISCUSSION

Dysregulated immune and salivary gland functions, including a prominent IFN signature, are implicated in the pathogenesis of SjD.^{12–70} Consistently, the whole blood RNA-seq from this study revealed IFN signalling dysregulation in the SjD^{Ro+}. In contrast, pathway analyses were indicative of T cell-driven disease mechanisms in the SjD^{Ro-}. Interestingly, the DE transcripts from both the SjD^{Ro+} and SjD^{Ro-} exhibited overall repression of transcript expression relative to HCs, including downregulation of many lncRNAs. Further, direct comparison of SjD^{Ro+} and SjD^{Ro-} yielded surprisingly few DE transcripts (online supplemental table 5), the majority of which were IFN-induced genes. Given previously reported differences in the absolute numbers of specific immune cell types in patients with SjD,¹⁶ the observed minor alterations in the abundance of immune cell types by deconvolution in this study (online supplemental figure

1), and the significant differences in the mean ages of the SjD and HC subjects from this study (online supplemental table 1), it is possible that these and other observations from this study may be influenced by the slight differences in the percentages of different immune cell types in the limited peripheral blood samples available from SjD cases subphenotyped based on Ro autoantibody status. Future studies using carefully matched case-control samples would be needed to resolve this limitation, as well as further improve our understanding of SjD heterogeneity.

Although recent studies have revealed important regulatory roles of several lncRNAs in immune responses and autoimmunity (reviewed in^{71–73}), few studies have investigated the potential roles of lncRNAs in SjD pathogenesis. The IFN signalling pathway is tightly regulated, requiring genetic ‘on/off’ switches to rapidly activate or suppress transcription in response to changing cellular environments.³³ More recent studies have identified lncRNAs that function as important regulators of IFN signalling, autoimmunity and cancer. Conflicting reports of *NRIR* functioning as a positive regulator of IFN-inducible gene expression in monocytes,^{26–27} but a negative regulator of expression in hepatocellular carcinomas,²⁷ suggest that *NRIR* expression is subject to cellular and context-specific regulatory effects. In this study, *NRIR* was overexpressed in whole blood of SjD^{Ro+} and induced by both type I and type II IFN stimulation in vitro. We also demonstrated that three previously uncharacterised antisense lncRNAs were IFN responsive: *OAS123-AS1*, *MX1-AS1* and *GBP5-AS1*. Our kinetic data suggest that these antisense lncRNAs are rapidly induced in response to IFNs and their induction may act as a massive upregulation signal to induce their respective pcRNAs. The precise mechanisms of action

for these novel antisense lncRNAs will require further studies.

We identified 15 other lncRNAs (online supplemental table 14) that were also correlated with IFN-inducible gene expression, including *BISPR* (BST2 IFN-stimulated positive regulator), which is implicated in the regulation of tetherin function,⁴⁰ and *CYTOR*, which interacts with microRNAs to modulate numerous signalling systems implicated in cancer.^{41–43} *CYTOR*, *BISPR*, *NRIR*⁷³ and *LINC00487*⁷⁴ (table 1) were previously reported as upregulated in patients with SjD. Interestingly, *LINC00487* resides ~50 kb upstream of *NRIR*, suggesting the entire genomic region containing *LINC00487*/*NRIR*/*CMK2*/*RSAD2* is subject to IFN regulation. However, other lncRNAs reportedly overexpressed in SjD, such as *IFNG-AS1*,³⁰ *NEAT1*²² and *PTVI*,²³ were not DE in our analyses (online supplemental tables 2–4). Further studies will be required to elucidate the mechanisms of action of undefined lncRNAs.

In this study, we identified *LINC01871* as overexpressed in all patients with SjD. Deep bioinformatics data mining, expression correlation analyses in the whole blood SjD^{Ro}-RNA-seq dataset, and pathway analyses all implicated *LINC01871* as a potential regulator of T cell activation and differentiation. Loss of *LINC01871* in a T cell line altered the expression of numerous transcripts implicated in innate and adaptive immune function. Dysregulation of *LINC01871* in SjD may play a role in the T cell-driven pathogenesis of SjD by impacting leucocyte migration and extravasation (*CD44*, *MMP2*, *MMP9*, *ITGAM*, *SELL*), IFN γ signalling (*CXCR3*) and cellular growth pathways (*CSF1*, *CSF2*).^{58–59–75} Notably, many of the pcRNAs perturbed due to *LINC01871* deletion have been previously implicated in the pathogenesis of SjD and/or autoimmunity. *MMP9* is reportedly overexpressed in the salivary gland of patients with SjD,^{63–64} and *CXCR3* is strongly induced by IFN γ and highly expressed in the T cells of patients with SjD.⁷⁵ *CSF2* is also reportedly elevated in the plasma of patients with SjD.⁷⁶ Further, *LINC01871* is overexpressed in several human cancers, highlighting the importance of this transcript in the regulation of multiple cellular processes.^{44–47}

Finally, we observed that *LINC01871* was negatively regulated by both short-term calcineurin/NFAT and TCR signalling in both a T cell line and primary human T cells, but positively regulated during long-term TCR stimulation. While it is tempting to assume an overall downregulation of *LINC01871* based on the use of PMA/I (figure 5A), this pharmacological stimulus is much more powerful and broadly acting than TCR signalling, modelled herein by cross-linking with anti-CD3/CD28 beads (figure 5F). The extent and duration of calcineurin activation after TCR signalling are distinct from that of PMA/I treatment, and this difference is readily appreciated in the bimodal regulation of *LINC01871* through longer stimulation and the effect of FK506 as an augmentor, rather than an inhibitor, of expression at late time points. These seemingly contradictory outcomes

point to complex regulation, likely by more than one pathway (eg, IFN γ , figure 4C). Further, we speculate that the upregulation of *LINC01871* during chronic or continual TCR stimulation is consistent with *LINC01871* dysregulation in response to dysregulated T cell function in autoimmune diseases. Dutta *et al* recently showed that calcineurin also has NFAT-independent activity by recruitment to the TCR and regulation of ICAM-1-dependent cell adhesion.⁷⁷ Our observed kinetic-dependent differences in *LINC01871* regulation in the primary human T cells emphasise the need for careful dissection of the involved molecular pathways in the future.

Our data clearly implicate the dysregulation of lncRNAs as an important mechanism driving the IFN-mediated and/or T cell-driven pathology of SjD. Understanding the underlying differential regulation of immunomodulatory gene expression in clinical subphenotypes of SjD will provide critical insights for future identification of biomarkers and/or development of novel therapeutic treatments. Further, the approaches described in this study provide a roadmap to guide future studies in the discovery and functional characterisation of novel lncRNAs with potential disease implications.

Data availability

Whole blood RNA-seq datasets generated during this study will be made available through the Databases of Genotypes and Phenotypes controlled-access study phs002723.v1.p1 after publication of this manuscript. RNA-seq datasets generated from the CRISPR-edited HSB-2 cell lines will be made available through the NCBI Sequence Read Archive after publication of this manuscript. Alternatively, access to the RNA-seq data will be made available in accordance with consent and data transfer/use agreement from the Oklahoma Medical Research Foundation by request to the corresponding author, Dr Christopher J Lessard (chris-lessard@omrf.org). All other data associated with this study were publicly available and cited in the manuscript and URLs provided as an online supplemental note.

Material availability

Participant whole blood RNAs used in this study were obtained from the Oklahoma Medical Research Foundation Sjögren's Research Clinic (OMRF-SRC) with Institutional Review Board (IRB) approval. Request for samples must be submitted to Dr A Darise Farris (principal investigator of OMRF-SRC; Darise-Farris@omrf.org). Access will depend on limited sample availability and will require IRB approval and completion of a material transfer agreement. *LINC01871*^{-/-} CRISPR-edited HSB-2 cell line generated in this study will be made available by request to the corresponding author (Dr Christopher Lessard; chris-lessard@omrf.org) after a material transfer agreement is completed. Sequences of the primers used to create and/or evaluate this cell line are provided in the online supplemental tables and can be ordered through

commercial sources. All other materials used in this study are commercially available.

Patient and public involvement

This study leveraged pre-existing banked whole blood RNA. Therefore, it was not appropriate or possible to involve patients or the public in the design, conduct, reporting or dissemination plans of our research.

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Contributors CJL supervised the research and had final responsibility for the decision to submit for publication. MLJ, BK, CL, JAI, AMS, NM, GBW, JDW and IA conceived, designed and/or performed the experiments and/or analyses under the supervision of CJL. KMG, JAK, DML, LR, DUS, JMG, JJ, RHAS, PMG, KS, AR, ADF and CJL collected and characterised the SjD cases and healthy controls used in this study. KMG manages the human sample and data curation. SBG manages the technology required for processing and analysis of large-scale sequencing data. JMG, JJ, PMG, CGM, KS, ADF, IA and CJL acquired funding that supported the research reported in this manuscript. All authors contributed intellectual content during the drafting and revision of the work and approved the final version to be published.

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Competing interests KS is a current employee of Janssen. ADF and CJL have an active collaborative research agreement with Janssen. All other authors have reported that they have no competing interests to report.

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the OMRF Sjögren's Research Clinic (OMRF-SRC) with institutional IRB Approval. Request for samples must be submitted to Dr. A. Darise Farris (PI of OMRF-SRC; darise-farris@omrf.org). Access will depend on limited sample availability and will require IRB Approval and completion of a material transfer agreement. LINC01871-/- CRISPR-edited HSB-2 cell lines generated in this study will be made available pending material transfer agreement upon request to Dr. Christopher J. Lessard. All other data associated with this study were publicly available and cited in this manuscript.

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REFERENCES

- Qin B, Wang J, Yang Z, *et al*. Epidemiology of primary Sjögren's syndrome: a systematic review and meta-analysis. *Ann Rheum Dis* 2015;74:1983–9.
- Helmick CG, Felson DT, Lawrence RC, *et al*. Estimates of the prevalence of arthritis and other rheumatic conditions in the United States. Part I. *Arthritis Rheum* 2008;58:15–25.
- Göransson LG, Haldorsen K, Brun JG, *et al*. The point prevalence of clinically relevant primary Sjögren's syndrome in two Norwegian counties. *Scand J Rheumatol* 2011;40:221–4.
- Goules AV, Kapsogeorgou EK, Tzioufas AG. Insight into pathogenesis of Sjögren's syndrome: dissection on autoimmune infiltrates and epithelial cells. *Clin Immunol* 2017;182:30–40.
- Mariette X, Criswell LA. Primary Sjögren's syndrome. *N Engl J Med* 2018;378:931–9.
- Imgenberg-Kreuz J, Rasmussen A, Sivils K, *et al*. Genetics and epigenetics in primary Sjögren's syndrome. *Rheumatology* 2021;60:2085–98.
- Björk A, Mofors J, Wahren-Herlenius M. Environmental factors in the pathogenesis of primary Sjögren's syndrome. *J Intern Med* 2020;287:475–92.
- Rasmussen A, Ice JA, Li H, *et al*. Comparison of the American-European consensus group Sjögren's syndrome classification criteria to newly proposed American College of rheumatology criteria in a large, carefully characterised sicca cohort. *Ann Rheum Dis* 2014;73:31–8.
- Shiboski SC, Shiboski CH, Criswell LA, *et al*. American College of rheumatology classification criteria for Sjögren's syndrome: a data-driven, expert consensus approach in the Sjögren's international collaborative clinical alliance cohort. *Arthritis Care Res* 2012;64:475–87.
- Vitali C, Bombardieri S, Jonsson R, *et al*. Classification criteria for Sjögren's syndrome: a revised version of the European criteria proposed by the American-European consensus group. *Ann Rheum Dis* 2002;61:554.
- Vitali C, Bootsma H, Bowman SJ, *et al*. Classification criteria for Sjögren's syndrome: we actually need to definitively resolve the long debate on the issue. *Ann Rheum Dis* 2013;72:476–8.
- Emamian ES, Leon JM, Lessard CJ, *et al*. Peripheral blood gene expression profiling in Sjögren's syndrome. *Genes Immun* 2009;10:285–96.
- Brito-Zerón P, Acar-Denizli N, Wan-Fai N, *et al*. How immunological profile drives clinical phenotype of primary Sjögren's syndrome at diagnosis: Analysis of 10,500 patients (Sjögren Big Data Project). *Clin Exp Rheumatol* 2018;36:S102–11.

- 14 Soret P, Le Dantec C, Desvaux E, *et al.* A new molecular classification to drive precision treatment strategies in primary Sjögren's syndrome. *Nat Commun* 2021;12:1–18.
- 15 Joachims ML, Leehan KM, Dozmorov MG, *et al.* Sjögren's syndrome minor Salivary Gland CD4+ memory T Cells associate with glandular disease features and have a germinal center T follicular helper transcriptional profile. *J Clin Med* 2020;9:2164–17.
- 16 Minguenau M, Boudaoud S, Haskett S, *et al.* Cytometry by time-of-flight immunophenotyping identifies a blood Sjögren's signature correlating with disease activity and glandular inflammation. *J Allergy Clin Immunol* 2016;137:e12:1809–21.
- 17 de Vita S, Gandolfo S, Zandonella Callegger S, *et al.* The evaluation of disease activity in Sjögren's syndrome based on the degree of MALT involvement: Glandular swelling and cryoglobulinaemia compared to ESSDAI in a cohort study. *Clin Exp Rheumatol* 2018;36:S150–6.
- 18 Engreitz JM, Ollikainen N, Guttman M. Long non-coding RNAs: spatial amplifiers that control nuclear structure and gene expression. *Nat Rev Mol Cell Biol* 2016;17:756–70.
- 19 Dolcino M, Tinazzi E, Vitali C, *et al.* Long non-coding RNAs modulate Sjögren's syndrome associated gene expression and are involved in the pathogenesis of the disease. *J Clin Med* 2019;8:1349.
- 20 Shi H, Cao N, Pu Y, *et al.* Long non-coding RNA expression profile in minor salivary gland of primary Sjögren's syndrome. *Arthritis Res Ther* 2016;18:1–14.
- 21 Sandhya P, Joshi K, Scaria V. Long noncoding RNAs could be potential key players in the pathophysiology of Sjögren's syndrome. *Int J Rheum Dis* 2015;18:898–905.
- 22 Ye L, Shi H, Yu C, *et al.* LncRNA NEAT1 positively regulates MAPK signaling and is involved in the pathogenesis of Sjögren's syndrome. *Int Immunopharmacol* 2020;88:106992.
- 23 Fu J, Shi H, Wang B, *et al.* LncRNA PVT1 links Myc to glycolytic metabolism upon CD4+ T cell activation and Sjögren's syndrome-like autoimmune response. *J Autoimmun* 2020;107:102358.
- 24 Cheng C, Zhou J, Chen R, *et al.* Predicted disease-specific immune infiltration patterns decode the potential mechanisms of long non-coding RNAs in primary Sjögren's syndrome. *Front Immunol* 2021;12.
- 25 Chen X, Cheng Q, Du Y, *et al.* Differential long non-coding RNA expression profile and function analysis in primary Sjögren's syndrome. *BMC Immunol* 2021;22.
- 26 Mariotti B, Servaas NH, Rossato M, *et al.* The long non-coding RNA NRIR drives IFN-Response in monocytes: implication for systemic sclerosis. *Front Immunol* 2019;10:100.
- 27 Kambara H, Niazi F, Kostadinova L, *et al.* Negative regulation of the interferon response by an interferon-induced long non-coding RNA. *Nucleic Acids Res* 2014;42:10668–80.
- 28 Dong G, Yang Y, Li X, *et al.* Granulocytic myeloid-derived suppressor cells contribute to IFN- γ signaling activation of B cells and disease progression through the lncRNA NEAT1-BAFF axis in systemic lupus erythematosus. *Biochim Biophys Acta Mol Basis Dis* 2020;1866:165554.
- 29 Pan Y, Wang T, Zhao Z, *et al.* Novel insights into the emerging role of NEAT1 and its effects downstream in the regulation of inflammation. *J Inflamm Res* 2022;15:557–71.
- 30 Wang J, Peng H, Tian J, *et al.* Upregulation of long noncoding RNA TM6VP1 enhances T helper type 1 cell response in patients with Sjögren syndrome. *Immunol Res* 2016;64:489–96.
- 31 Loureiro-Amigo J, Palacio-García C, Martínez-Gallo M, *et al.* Utility of lymphocyte phenotype profile to differentiate primary Sjögren's syndrome from sicca syndrome. *Rheumatology* 2021;60:5647–58.
- 32 Plattner C, Finotello F, Rieder D. Deconvoluting tumor-infiltrating immune cells from RNA-Seq data using quanTIseq. *Methods Enzymol* 2020;636:261–85.
- 33 Muskardin TLW, Niewold TB. Type I interferon in rheumatic diseases. *Nat Rev Rheumatol* 2018;14:214–28.
- 34 Subramanian A, Tamayo P, Mootha VK, *et al.* Gene set enrichment analysis: a knowledge-based approach for interpreting genome-wide expression profiles. *Proc Natl Acad Sci U S A* 2005;102:15545–50.
- 35 Subramanian A, Kuehn H, Gould J, *et al.* GSEA-P: a desktop application for gene set enrichment analysis. *Bioinformatics* 2007;23:3251–3.
- 36 Rinchai D, Roelands J, Toufiq M, *et al.* BloodGen3Module: blood transcriptional module repertoire analysis and visualization using R. *Bioinformatics* 2021;37:2382–9.
- 37 Li S, Roupahel N, Duraisingham S, *et al.* Molecular signatures of antibody responses derived from a systems biology study of five human vaccines. *Nat Immunol* 2014;15:195–204.
- 38 Chaussabel D, Quinn C, Shen J, *et al.* A modular analysis framework for blood genomics studies: application to systemic lupus erythematosus. *Immunity* 2008;29:150–64.
- 39 Caielli S, Cardenas J, de Jesus AA, *et al.* Erythroid mitochondrial retention triggers myeloid-dependent type I interferon in human SLE. *Cell* 2021;184:e19:4464–79.
- 40 Barriocanal M, Carrero E, Segura V, *et al.* Long non-coding RNA BST2/BISPR is induced by IFN and regulates the expression of the antiviral factor tetherin. *Front Immunol* 2014;5:655.
- 41 Liang J, Wei X, Liu Z, *et al.* Long noncoding RNA CYTOR in cancer: a TCGA data review. *Clin Chim Acta* 2018;483:227–33.
- 42 Yang J, Ma Q, Zhang M, *et al.* LncRNA CYTOR drives L-OHP resistance and facilitates the epithelial-mesenchymal transition of colon carcinoma cells via modulating miR-378a-5p/SERPINE1. *Cell Cycle* 2021;20:1415–30.
- 43 Chen W, Du M, Hu X, *et al.* Long noncoding RNA cytoskeleton regulator RNA promotes cell invasion and metastasis by titrating miR-613 to regulate ANXA2 in nasopharyngeal carcinoma. *Cancer Med* 2020;9:1209–19.
- 44 Chen Q, Hu L, Huang D, *et al.* Six-lncRNA immune prognostic signature for cervical cancer. *Front Genet* 2020;11:1252.
- 45 He Y, Wang X, Biomedical W X. Identification of molecular features correlating with tumor immunity in gastric cancer by multi-omics data analysis. *Ann Transl Med* 2020;8:1050.
- 46 Feng Y, Shen Y, Chen H, *et al.* Expression profile analysis of long non-coding RNA in acute myeloid leukemia by microarray and bioinformatics. *Cancer Sci* 2018;109:340–53.
- 47 Ma W, Zhao F, Yu X, *et al.* Immune-related lncRNAs as predictors of survival in breast cancer: a prognostic signature. *J Transl Med* 2020;18:1–13.
- 48 Xu M, Zhang R, Qiu J. A four immune-related long noncoding RNAs signature as predictors for cervical cancer. *Hum Cell* 2022;35:348–59.
- 49 Wang Z, Zhang J, Liu Y, *et al.* An integrated autophagy-related long noncoding RNA signature as a prognostic biomarker for human endometrial cancer: a bioinformatics-based approach. *Biomed Res Int* 2020;2020:5717498.
- 50 Volders P-J, Anckaert J, Verheggen K, *et al.* LNCipedia 5: towards a reference set of human long non-coding RNAs. *Nucleic Acids Res* 2019;47:D135–9.
- 51 Huang N, Pérez P, Kato T, *et al.* SARS-CoV-2 infection of the oral cavity and saliva. *Nat Med* 2021;27:892–903.
- 52 Yin H, Pranzatelli TJF, French BN, *et al.* Sclerosing Sialadenitis is associated with salivary gland hypofunction and a unique gene expression profile in Sjögren's syndrome. *Front Immunol* 2021;12.
- 53 van Dam S, Craig T, de Magalhães JP. GeneFriends: a human RNA-seq-based gene and transcript co-expression database. *Nucleic Acids Res* 2015;43:D1124–32.
- 54 Perron U, Provero P, Molineris I. In silico prediction of lncRNA function using tissue specific and evolutionary conserved expression. *BMC Bioinformatics* 2017;18:144.
- 55 Jiang Q, Ma R, Wang J, *et al.* LncRNA2Function: a comprehensive resource for functional investigation of human lncRNAs based on RNA-Seq data. *BMC Genomics* 2015;16 Suppl 3:S2.
- 56 Park Y-J, Yoo S-A, Kim M, *et al.* The role of calcium-calciurein-NFAT signaling pathway in health and autoimmune diseases. *Front Immunol* 2020;11:195.
- 57 CarbonS, DouglassE, DunnN, *et al.* The gene ontology resource: 20 years and still going strong. *Nucleic Acids Res* 2019;47:D330–8.
- 58 Fuhrman CA, Yeh W-I, Seay HR, *et al.* Divergent phenotypes of human regulatory T cells expressing the receptors TIGIT and CD226. *J Immunol* 2015;195:145–55.
- 59 Deng C, Chen Y, Li W, *et al.* Alteration of CD226/TIGIT immune checkpoint on T cells in the pathogenesis of primary Sjögren's syndrome. *J Autoimmun* 2020;113:102485.
- 60 Miyagawa I, Nakayama S, Nakano K, *et al.* Induction of regulatory T cells and its regulation with insulin-like growth factor/insulin-like growth factor binding protein-4 by human mesenchymal stem cells. *J Immunol* 2017;199:1616–25.
- 61 Merad M, Sathe P, Helft J, *et al.* The dendritic cell lineage: ontogeny and function of dendritic cells and their subsets in the steady state and the inflamed setting. *Annu Rev Immunol* 2013;31:563–604.
- 62 Satpathy AT, Wu X, Albring JC, *et al.* Re(defined) the dendritic cell lineage. *Nat Immunol* 2012;13:1145–54.
- 63 Pérez P, Kwon Y-J, Allende C, *et al.* Increased acinar damage of salivary glands of patients with Sjögren's syndrome is paralleled by simultaneous imbalance of matrix metalloproteinase 3/tissue inhibitor of metalloproteinases 1 and matrix metalloproteinase 9/tissue inhibitor of metalloproteinases 1 ratios. *Arthritis Rheum* 2005;52:2751–60.
- 64 Asatsuma M, Ito S, Watanabe M, *et al.* Increase in the ratio of matrix metalloproteinase-9 to tissue inhibitor of metalloproteinase-1 in saliva from patients with primary Sjögren's syndrome. *Clinica Chimica Acta* 2004;345:99–104.

- 65 Witkowska AM. Soluble ICAM-1: a marker of vascular inflammation and lifestyle. *Cytokine* 2005;31:127–34.
- 66 Gaud G, Lesourne R, Love PE. Regulatory mechanisms in T cell receptor signalling. *Nat Rev Immunol* 2018;18:485–97.
- 67 Burnett RC, Thirman MJ, Rowley JD, *et al.* Molecular analysis of the T-cell acute lymphoblastic leukemia-associated t(1;7)(p34;q34) that fuses LCK and TCRB. *Blood* 1994;84:1232–6.
- 68 Annett S, Moore G, Robson T. FK506 binding proteins and inflammation related signalling pathways; basic biology, current status and future prospects for pharmacological intervention. *Pharmacol Ther* 2020;215:107623.
- 69 Ehx G, Somja J, Warnatz H-J, *et al.* Xenogeneic graft-versus-host disease in humanized NSG and NSG-HLA-A2/HHD mice. *Front Immunol* 2018;9:1943.
- 70 Hjelmervik TOR, Petersen K, Jonassen I, *et al.* Gene expression profiling of minor salivary glands clearly distinguishes primary Sjögren's syndrome patients from healthy control subjects. *Arthritis Rheum* 2005;52:1534–44.
- 71 Zou Y, Xu H. Involvement of long noncoding RNAs in the pathogenesis of autoimmune diseases. *J Transl Autoimmun* 2020;3:100044.
- 72 De Benedittis G, Ciccacci C, Latini A, *et al.* Emerging role of microRNAs and long non-coding RNAs in Sjögren's syndrome. *Genes* 2021;12. doi:10.3390/genes12060903. [Epub ahead of print: 11 06 2021].
- 73 Peng Y, Luo X, Chen Y, *et al.* LncRNA and mRNA expression profile of peripheral blood mononuclear cells in primary Sjögren's syndrome patients. *Sci Rep* 2020;10.
- 74 Inamo J, Suzuki K, Takeshita M, *et al.* Identification of novel genes associated with dysregulation of B cells in patients with primary Sjögren's syndrome. *Arthritis Res Ther* 2020;22:153.
- 75 Yao Y, Ma J-F, Chang C, *et al.* Immunobiology of T cells in Sjögren's syndrome. *Clin Rev Allergy Immunol* 2021;60:111–31.
- 76 Greenwell-Wild T, Moutsopoulos NM, Gliozzi M, *et al.* Chitinases in the salivary glands and circulation of patients with Sjögren's syndrome: macrophage harbingers of disease severity. *Arthritis Rheum* 2011;63:3103–15.
- 77 Dutta D, Barr VA, Akpan I, *et al.* Recruitment of calcineurin to the TCR positively regulates T cell activation. *Nat Immunol* 2017;18:196–204.